

**Dietary fiber intervention on gut microbiota composition in healthy adults: A systematic review and meta-analysis**

So, Daniel; Whelan, Kevin; Rossi, Megan; Morrison, Mark; Holtmann, Gerald; Kelly, Jaimon T.; Shanahan, Erin R.; Staudacher, Heidi M.; Campbell, Katrina L.

*Published in:*  
American Journal of Clinical Nutrition

*DOI:*  
[10.1093/ajcn/nqy041](https://doi.org/10.1093/ajcn/nqy041)

Published: 01/06/2018

*Document Version:*  
Peer reviewed version

[Link to publication in Bond University research repository.](#)

*Recommended citation(APA):*

So, D., Whelan, K., Rossi, M., Morrison, M., Holtmann, G., Kelly, J. T., Shanahan, E. R., Staudacher, H. M., & Campbell, K. L. (2018). Dietary fiber intervention on gut microbiota composition in healthy adults: A systematic review and meta-analysis. *American Journal of Clinical Nutrition*, 107(6), 965-983.  
<https://doi.org/10.1093/ajcn/nqy041>

**General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

For more information, or if you believe that this document breaches copyright, please contact the Bond University research repository coordinator.

# **Dietary fiber intervention on gut microbiota composition in healthy adults: a systematic review and meta-analysis**

## **Review authors**

Daniel So, APD

Kevin Whelan, PhD

Megan Rossi, APD, PhD

Mark Morrison, PhD

Gerald Holtmann, PhD

Jaimon T. Kelly, APD, PhD Candidate

Erin R Shanahan, PhD

Heidi M Staudacher, APD, PhD

Katrina L. Campbell, AdvAPD, PhD

## **Affiliations**

1. Faculty of Health Sciences and Medicine, Bond University, Gold Coast, Australia (DS; JK; KC)
2. King's College, London, Department of Nutritional Sciences, United Kingdom (MR; KW)
3. The University of Queensland Diamantina Institute, The University of Queensland, Translational Research Institute, Brisbane, Australia (ES; MM)
4. Faculty of Medicine, University of Queensland, Brisbane, Australia (MM, GH, HS)
5. Department of Gastroenterology & Hepatology, Princess Alexandra Hospital, Brisbane, Australia (GH, ES)
6. Department of Nutrition and Dietetics, Princess Alexandra Hospital, Brisbane, Australia (KC)

## **Authors last names**

So, Whelan, Rossi, Morrison, Holtmann, Kelly, Shanahan, Staudacher, Campbell

## **Disclaimers**

None.

## **Corresponding author**

Katrina L Campbell, [kcampbel@bond.edu.au](mailto:kcampbel@bond.edu.au)

Faculty of Health Science and Medicine, Bond University

14 University Drive, Robina, Queensland, 4226, Australia

Phone: (07) 559 53573

## **Sources of support**

This work has received no specific funding.

## **Short running head**

Dietary fiber interventions on the gut microbiota

## **Abbreviations**

CI – Confidence intervals

FISH – Fluorescence *in situ* hybridization

GI – Gastrointestinal

HMO – Human Milk Oligosaccharide

ICTRP – International Clinical Trials Register

MD – Mean difference

OTU – Operational taxonomic unit

PRISMA – Preferred Reporting Items for Systematic Reviews and Meta-Analysis

PROSPERO – The International Prospective Register of Systematic Reviews

qPCR – Quantitative polymerase chain reaction

RCT – Randomized controlled trial

SCFA – Short chain fatty acid

SD – Standard deviation

SE – Standard error

SMD – Standardized mean difference

**Clinical trial registry number**

Not required. PROSPERO registration (CRD42016053101)

URL: [http://www.crd.york.ac.uk/PROSPERO/display\\_record.asp?ID=CRD42016053101](http://www.crd.york.ac.uk/PROSPERO/display_record.asp?ID=CRD42016053101)

## 1 ABSTRACT

2 **Background:** Dysfunction of the gut microbiota is frequently reported as a manifestation of  
3 chronic disease, and therefore presents as a modifiable risk factor in their development. Diet is  
4 a major regulator of the gut microbiota and certain types of dietary fiber may modify bacterial  
5 numbers and metabolism, including short-chain fatty acid (SCFA) generation.

6 **Objective:** A systematic review and meta-analysis were undertaken to assess the effect of  
7 dietary fiber interventions on gut microbiota composition in healthy adults.

8 **Design:** A systematic search was conducted across MEDLINE, EMBASE, CENTRAL and  
9 CINAHL for randomized controlled trials using culture and/or molecular microbiological  
10 techniques evaluating the effect of fiber intervention on gut microbiota composition in healthy  
11 adults. Meta-analyses using random-effects model were performed on alpha diversity, pre-  
12 specified bacterial abundances including *Bifidobacterium* and *Lactobacillus* spp., and fecal  
13 SCFA concentrations comparing dietary fiber intervention with placebo/low fiber  
14 comparators.

15 **Results:** A total of 64 studies involving 2099 participants were included. Dietary fiber  
16 intervention resulted in higher abundance of *Bifidobacterium* spp. [Standardized Mean  
17 Difference (SMD) 0.64 (95% Confidence Interval: 0.42, 0.86];  $P < 0.00001$ ] and *Lactobacillus*  
18 spp. [SMD: 0.22 (0.03, 0.41),  $P = 0.02$ ] as well as fecal butyrate concentration [SMD: 0.24  
19 (0.00, 0.47),  $P = 0.05$ ] compared with placebo/low fiber comparators. Subgroup analysis  
20 revealed fructans and galacto-oligosaccharides led to significantly greater abundance of both  
21 *Bifidobacterium* spp. and *Lactobacillus* spp. compared with comparators ( $P < 0.00001$  and  $P =$   
22 0.002 respectively). No differences in effect were found between fiber intervention and  
23 comparators for  $\alpha$ -diversity, abundances of other pre-specified bacteria, or other SCFA  
24 concentrations.

25 **Conclusion:** Dietary fiber intervention, particularly involving prebiotic fibers, leads to higher  
26 fecal abundance of *Bifidobacterium* and *Lactobacillus* spp. but does not impact  $\alpha$ -diversity.  
27 Further research is required to better understand the role of individual fiber types on the  
28 growth of microbes and the overall gut microbial community.

29 **KEYWORDS**

30 Diet, dietary fiber, gastrointestinal microbiome, gastrointestinal microbiota, gut microbiota,  
31 prebiotic

## BACKGROUND

The gut microbiota is a highly diverse and metabolically active community, consisting of approximately  $3.9 \times 10^{13}$  microbial cells (1). These microbes participate in several functions beneficial to the host, including the fermentation of undigested nutrients (2, 3), synthesis of vitamins (4) and interaction with the immune system (5, 6). A number of disorders, including irritable bowel syndrome and type 2 diabetes mellitus, have been linked with disturbances in gut microbiota composition (2, 7-9). Such an association presents the gut microbiota as a potentially modifiable risk factor in the etiology of these conditions.

The gut microbiota can be detected and enumerated using different methods ranging from culture to next-generation sequencing (6, 10, 11), and can be characterized by measures of diversity and bacterial abundances (12, 13). Alpha diversity of the gut microbiota describes the richness (number of taxonomically distinct organisms present) and evenness (relative abundances of organisms) of its composition (12, 13), with cross-sectional studies demonstrating inverse associations between  $\alpha$ -diversity and disease states (7-9). Specific bacteria shown to be more abundant in health compared with disease states include *Bifidobacterium* and *Lactobacillus* spp. (2, 7, 14), whose functions include carbohydrate fermentation and vitamin synthesis (15-18). Furthermore, increasing evidence supports the importance of 'keystone' bacterial species, whose absence may have profound consequences for the host, as well as other members of the microbial community and their metabolic outputs, including the short-chain fatty acid (SCFA) butyrate (19-23). Butyrate is of particular interest to health due to its beneficial properties such as its immunomodulatory effects (24, 25).

Dietary fiber is defined as non-digestible carbohydrates of  $\geq 3$  monomeric units found inherently in foods, and also includes isolated or synthetic fibers with demonstrated physiological benefits (26-28). It is a key candidate in facilitating changes in the gut microbiota, as it escapes digestion by the host in the small intestine to pass into the colon

where it is available to the microbial community. Dietary fiber encompasses an array of heterogeneous compounds whose physicochemical properties vary based on their particle size, chemical structure, solubility, viscosity and fermentability (29, 30). Fiber with fermentable characteristics are substrates for the microbial population in the colon, stimulating growth of specific organisms and leading to production of various metabolites including SCFA (19, 29, 31). Indeed, some fibers can be further classified as ‘prebiotic’ (e.g. fructans) if they have been shown to be selectively utilized by host microorganisms conferring a health benefit (32). The current body of evidence regarding the effect of dietary fiber on the gut microbiota is informed via specific prebiotic fiber interventions (33, 34), whole-diet interventions (35-37) and cross-sectional associations (38, 39). However, these investigations are limited in that prebiotic fibers represent only a subset of total dietary fiber, and confounding factors such as disease states and intake of other fermentable substrates, are unaccounted for in whole diet studies and cross-sectional studies (40). Therefore, there is a gap in knowledge regarding the precise impact of dietary fiber intervention on the gut microbiota in healthy subjects, and this is the focus on the systematic review.

## **METHODS**

This systematic review was conducted in line with the guidelines of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis: The PRISMA statement (41), and the guidelines of the Cochrane Handbook for Systematic Reviews and Interventions (42). The methods including the eligibility criteria, search strategy, extraction process and analysis were pre-specified and documented in a protocol that was published in the International Prospective Register of Systematic Reviews (CRD42016053101).

### **Literature search**

A literature search was performed in the electronic databases MEDLINE, EMBASE, CENTRAL and CINAHL (from inception to October 4, 2017), using a combination of subject



headings, free text terms and synonyms relevant to this review, in consultation with an experienced systematic review search librarian (**Supplemental Tables 1-4**). There was no date or language restriction in the search strategy. A multi-step search approach was taken to retrieve relevant studies through additional hand-searching; contacting field experts; searching conference abstracts; theses and dissertations (ProQuest); and the International Clinical Trials Register (ICTRP) Search Portal and ClinicalTrials.gov to identify ongoing trials. Two review authors (DS and HS) screened articles in a blinded, standardized manner, with disagreements in judgement resolved by consensus or a third reviewer (KC).

## **Study selection**

Search results were merged into reference management software Endnote (X7; Thomson Reuters) and de-duplicated prior to screening using Rayyan (Qatar Computing Research Institute) (43). Full text articles of potentially relevant studies were sought and reviewed. Attempts were made to contact the corresponding author where the full text article provided inadequate information to assess eligibility or extract relevant data. Studies were included if they met all of the following criteria: 1) randomized controlled trial (RCT), cluster RCT, or quasi-RCT; 2) inclusion of healthy adult participants ( $\geq 18$  years of age); 3) intervention aimed at increasing fiber intake; 4) inclusion of a placebo for supplement interventions (e.g. maltodextrin), and either low fiber control (e.g. white bread) or habitual diet group for food interventions as comparators; 5) measured fecal microbiota related outcomes at the end of intervention.

Studies that were solely investigating enteral nutrition and those that included participants with an acute or chronic disease, including gastrointestinal (GI) conditions such as coeliac disease, inflammatory bowel disease, irritable bowel syndrome and other functional gastrointestinal disorders were excluded. Studies including mixed population groups where the healthy subgroup was not reported separately were also excluded. Studies that included overweight and

obese participants who were otherwise healthy and without any abnormal clinical parameters (e.g. elevated blood pressure) were included. Interventions eligible for inclusion provided an increase in fiber intake achieved through 1) dietary counselling to increase dietary fiber intake from food 2) food intervention (e.g. added cereals); or 3) fiber supplementation. Dietary counselling studies or food interventions were only included if fiber modification was the primary aim of the intervention.

The primary outcome was between-group differences in  $\alpha$ -diversity of fecal microbiota at the end of the intervention. Measures of  $\alpha$ -diversity included the total number of observed operational taxonomic units (OTUs) (the number of taxonomically-related groups of bacteria, evaluating richness); Chao1 Index (a non-parametric richness estimator); Shannon diversity index (a metric combining richness and evenness, with equal weighting to abundant and rare species); and Simpson diversity index (metric of richness and evenness, where more weighting is given to abundant species). Secondary outcomes were between-group differences in abundances of the following commonly measured bacterial groups: *Bifidobacterium* spp.; *Lactobacillus* spp.; *Roseburia* spp.; *Akkermansia muciniphila*; *Eubacterium hallii*; *Eubacterium rectale*; *Faecalibacterium prausnitzii*; and *Ruminococcus bromii*. Studies were included if they reported on either primary or secondary outcomes. Between-group differences in fecal SCFAs (total SCFAs and butyrate) were included as an exploratory outcome.

### **Data extraction and management**

Two reviewers (DS and HS) independently extracted the data from eligible studies. Data extracted included: study design (duration, location, details of ‘run-in’ and ‘wash-out’ periods); participant characteristics, intervention details (fiber type, fiber dose, intervention delivery, compliance, assessment and control of dietary intake); and other information including antibiotic or probiotic use.

For all pre-specified primary, secondary and exploratory outcome data, the mean, standard deviation (SD), standard error (SE) or 95% confidence intervals (CI) that were reported at end of intervention were extracted for analysis. Where studies used multiple intervention groups of different fiber doses, data for the highest intervention dose was extracted. Where studies used multiple intervention groups of different fibers at the same dose compared with a single control group, data was extracted from each intervention group and pooled together. A weighted average of the intervention groups and the study variance was then calculated (44). Risk of bias was independently assessed by two reviewers (DS and HS) using Cochrane methodology (45). The review assessed “other bias” regarding the control of dietary intake during the study. This included examining whether dietary advice (e.g. to maintain dietary intake or avoid probiotic food sources) was provided, whether dietary compliance and/or intake were measured and reported, and if adjustments in statistical analysis were made if differences in dietary intake were found.

#### **Statistical analysis**

The overall treatment effect of fiber on primary and secondary outcomes was calculated using the difference between the end of intervention values for the intervention and comparator groups. Data reported as median and interquartile range were converted to mean and SD as previously described (46). Variance was calculated from the SD and SE of end of intervention values, or from the confidence intervals (CI) where these values were not available (46). In crossover studies, the mean and SD, SE or CI of intervention and control periods were extracted and analyzed separately (47). Where end of intervention endpoint data was unable to be obtained, the results were described in text only.

Meta-analysis was performed where outcomes were reported in at least two studies using Revman (Version 5.3; Cochrane Collaboration). The mean difference (MD) was used to calculate effect sizes where outcome data were presented in the same units (Shannon diversity

index, total number of observed OTUs). Where outcome data were reported using different units, effect sizes were calculated using the standardized mean difference (SMD) (bacterial abundances, fecal SCFA concentration).

A random-effects model was used to produce a pooled estimate of the MD or SMD, and the fixed-effects model was used to check for robustness and potential outliers. Inconsistencies between studies were assessed using the  $I^2$  statistic, where significant heterogeneity was defined as  $I^2 \geq 50\%$ .

Pre-defined subgroup analyses were undertaken for primary and secondary outcomes that were reported in at least two studies in each subgroup. Pre-defined subgroup analyses included intervention types (supplements and dietary interventions), fiber types (accepted and candidate prebiotic fibers defined by Roberfroid et al., and general fibers defined by the review) (34), dose-response (comparing difference in fiber intake between intervention and control group of  $\leq 5\text{g/d}$ ,  $5\text{-}10\text{g/d}$ , and  $>10\text{g/d}$ ), trial design (parallel and crossover), and microbial analysis method (e.g. culture, sequencing). Post hoc subgroup analyses were undertaken for exploratory outcomes based on reporting method of fecal SCFA concentrations (dry weight of feces and wet weight of feces). Fructans and galacto-oligosaccharides were classified as ‘accepted prebiotic’ fibers, while ‘candidate prebiotic’ fibers included a broader range of fibers including polydextrose and resistant starch (34). The term ‘general fiber’ was used by the review to describe fibers not classified as either accepted or candidate prebiotics, and is not a formal term used to describe fibers in the literature.

For the fiber type subgroup analysis only, the fiber arm with the highest prebiotic classification (e.g. accepted prebiotic as opposed to a general fiber) was selected if multiple intervention groups were reported. Where multiple arms of the same prebiotic classification were presented, the interventions were pooled together and a weighted average of the intervention arms and study variance were calculated (44). Significant outliers were determined by visual

inspection as well as through a study-by-study sensitivity analysis, where each study was sequentially omitted and the remaining data re-assessed. If a study contributed to over 30% heterogeneity (based on changes to the  $I^2$  statistic) then it was removed from the analysis in the sensitivity analysis. Funnel plots were generated for outcomes where at least 10 studies were included in meta-analysis (48) and reporting bias detected by assessment of funnel plot asymmetry by visual inspection.

## RESULTS

### Study characteristics

Study identification and selection are detailed in the PRISMA flow chart (**Figure 1**). The initial electronic and manual search generated 3829 records. After review of full texts (**Supplemental Table 5**), 64 publications, along with three secondary studies (49-51) reporting additional outcomes from the primary publications, fulfilled the inclusion criteria and were included in the review.

The 64 included primary studies that analyzed a total of 2099 participants. Of these 64 studies, 29 were parallel RCTs (52-80) and 35 were crossover RCTs (81-115). Five crossover trials did not include a wash out period (84, 93, 95, 105, 108). The majority of studies (52 studies) used fiber supplementation, including: accepted prebiotic fiber (26 studies) (52, 54-58, 61, 62, 65, 67, 70, 74, 86, 90, 92, 95, 97, 100, 102, 103, 105, 107, 109-111, 115); candidate prebiotic fiber (18 studies) (53, 63, 64, 66, 68, 69, 73, 77, 81, 83, 84, 87, 88, 91, 99, 101, 112, 113); general fiber (seven studies) (59, 60, 72, 76, 80, 93, 94); and a fiber mix (108). The remaining 12 studies used food intervention by providing key food items (e.g. wholegrain cereal) to supplement the diet (71, 78, 82, 85, 89, 96, 98) or provided all food and fluid to participants (75, 79, 104, 106, 114). Intervention doses ranged from 1.2 g/d to 50 g/d, while treatment periods ranged from five days to three months, with a median length of three weeks.

Analysis techniques used to characterize fecal microbiota included: culture (15 studies) (52, 54-58, 65, 66, 69, 71, 73, 96, 98, 105, 114); fluorescence *in situ* hybridization (FISH) (20 studies) (53, 70, 74, 76, 82, 85, 89-92, 94, 99, 100, 103, 106, 108-110, 112, 113); quantitative polymerase chain reaction (qPCR) (11 studies) (60, 63, 68, 81, 86, 87, 95, 102, 104, 107, 111); and next-generation sequencing (including 454 pyrosequencing and Illumina sequencing) (12 studies) (59, 62, 64, 72, 75, 77-80, 97, 101, 115). A combination of techniques were used in six studies (49, 61, 67, 83, 84, 88, 93).

The outcomes of each meta-analysis are reported in **Table 1**. Results from subgroup analyses performed are included in **Supplemental Table 6**. Overall, outcome data from 56 studies were suitable for meta-analysis; results from the following studies were unable to be statistically pooled and are presented narratively under their respective sub-headings (59, 62, 69, 77-79, 83, 93, 95, 97, 101, 113, 115). The characteristics of included studies are presented in **Tables 2-3**.

#### **Dietary fiber and gut microbiota diversity ( $\alpha$ -diversity)**

Alpha-diversity was measured in 13 studies involving 393 participants (49, 59, 64, 72, 75, 77, 79, 80, 83, 88, 93, 97, 101).

Ten studies reported  $\alpha$ -diversity using Shannon diversity index. Of these, six reported the metric in a form suitable for inclusion in the meta-analysis (49, 64, 72, 75, 80, 88). Dietary fiber intervention had no effect on  $\alpha$ -diversity compared with placebo/low fiber comparators [MD: -0.06 Shannon diversity index (95% CI: -0.25, 0.12),  $P = 0.48$ ], albeit with substantial heterogeneity ( $I^2 = 53\%$ ). In two of the studies not included in the meta-analysis, raffinose and resistant starch interventions did not lead to significant difference in  $\alpha$ -diversity compared with placebo (93, 101). A significant reduction in the  $\alpha$ -diversity of fecal microbiota from baseline was detected in a trial involving flaxseed mucilage, measured by both the exponential of Shannon diversity index [-38010 (95% CI: -64473, -11546,  $P = 0.007$ )] as well as through

Simpson's inverse index [-17515 (95% CI: -30992, -4038,  $P = 0.014$ )], although a between-group comparison was not reported (59). Conversely, significant end of intervention differences in  $\alpha$ -diversity measured by Shannon diversity index ( $P = 0.013$ ) and inverse Simpson index ( $P = 0.004$ ) were detected between intervention and comparator groups in a supplementation trial involving resistant starch type 2 (77).

A study evaluating  $\alpha$ -diversity through Simpson's index found it was significantly higher in the intervention group receiving polydextrose compared with placebo after 21 days ( $P = 0.014$ ) (88). A trial involving 15 g/d arabinoxylan supplementation reported variable intervention effects when  $\alpha$ -diversity was evaluated using different metrics:  $\alpha$ -diversity was significantly lower compared with placebo when measured through observed species ( $P = 0.029$ ), but there were no significant differences when assessed by Simpson's evenness ( $P = 0.063$ ) (80).

A separate meta-analysis was performed for the three studies reporting  $\alpha$ -diversity measured by total number of observed OTUs (49, 72, 75). Dietary fiber had no effect on  $\alpha$ -diversity compared with placebo/low fiber comparators [MD: -4.37 OTUs (95% CI: -42.92, 34.19),  $P = 0.82$ ], with no heterogeneity ( $I^2 = 0\%$ ). The Chao1 index was used to report  $\alpha$ -diversity in two studies, although there was insufficient data available precluding meta-analysis. Neither trial reported significant differences between fiber intervention and placebo or low fiber control (49, 83). A feeding trial comparing wholegrain and refined grain diets found no difference in  $\alpha$ -diversity at end of intervention between the two groups, although the metric used to measure  $\alpha$ -diversity was not reported (79).

### **Dietary fiber and bacterial abundances**

Reporting of bacterial abundances differed across studies. Of the taxa of interest in this review, abundances of *Bifidobacterium* spp. (59 studies) and *Lactobacillus* spp. (28 studies) were most commonly reported. No studies reported on the abundance of *Akkermansia muciniphila*.

A total of 59 studies including 1896 participants reported the effect of dietary fiber on *Bifidobacterium* spp. abundance and of these, 51 trials (1629 participants) reported data in a form suitable for meta-analysis (53-58, 60, 61, 63-68, 70, 71, 73-76, 81, 82, 84-94, 96-112, 114). Dietary fiber led to a significantly greater *Bifidobacterium* spp. abundance compared with placebo/low fiber comparators [SMD: 0.64 (95% CI: 0.42, 0.86),  $P < 0.00001$ ], albeit with considerable heterogeneity ( $I^2 = 85\%$ ) (**Figure 2**).

However, subgroup analysis showed fiber interventions delivered through supplements resulted in a significantly higher *Bifidobacterium* spp. abundance compared with placebo/low fiber controls [SMD: 0.75 (95% CI: 0.52, 0.98),  $P < 0.00001$ ,  $I^2 = 83\%$ ], whereas no differences were found between food interventions and comparators [SMD: 0.20 (95% CI: -0.36, 0.76),  $P = 0.49$ ,  $I^2 = 88\%$ ], although considerable heterogeneity persisted in both analyses.

Subgroup analysis demonstrated interventions investigating fibers classified as accepted prebiotics and candidate prebiotics resulted in a significantly higher *Bifidobacterium* spp. abundance compared with placebo/low fiber controls [Accepted prebiotic fiber SMD: 0.68 (95% CI: 0.38, 0.98),  $P < 0.00001$ ,  $I^2 = 81\%$ ; Candidate prebiotic fiber SMD: 0.77 (95% CI: 0.30, 1.24),  $P < 0.00001$ ,  $I^2 = 86\%$ ] (**Figure 2**). However, there was no difference in effect between the general fiber subgroup compared with comparators [SMD: 0.25 (95% CI: -0.16, 0.65),  $P = 0.24$ ,  $I^2 = 86\%$ ]. This subgroup analysis did not reduce the considerable heterogeneity across each subgroup.

Subgroup analysis of dose-response showed dietary fiber led to significantly higher *Bifidobacterium* spp. abundance compared with placebo/low fiber comparators at all pre-defined dosage [ $\leq 5$ g/d fiber SMD: 0.51 (95% CI: 0.18, 0.84),  $P = 0.003$ ,  $I^2 = 70\%$ ; 5-10g/d SMD: 0.48 (95% CI: 0.13, 0.83),  $P = 0.007$ ,  $I^2 = 87\%$ ;  $>10$ g/d SMD: 0.85 (95% CI: 0.45, 1.25),



279  $P < 0.00001$ ,  $I^2 = 85\%$ ]. No differences were found in subgroup analyses of trial design or  
 280 microbiota analysis method (**Supplemental Table 6**).  
 281 Eight trials were not included in the meta-analysis. In the supplement trials of accepted  
 282 prebiotics, a significantly higher *Bifidobacterium* spp. abundance was reported following  
 283 supplementation involving inulin (115) and human milk oligosaccharides (HMO) (62)  
 284 compared with placebo at the end of intervention, while a significant within-group increase  
 285 from baseline was detected following 10g/d inulin supplementation (95). In the candidate  
 286 prebiotic trial of resistant starch supplementation, *Bifidobacterium* spp. abundance was  
 287 significantly higher in the intervention group compared with placebo at end of intervention  
 288 (77). In the supplement studies of general fiber, *Bifidobacterium* spp. abundance was higher  
 289 following after xylo-oligosaccharide supplementation compared with placebo (69) while  
 290 manno-oligosaccharides had no effect on *Bifidobacterium* spp. compared with placebo (113).  
 291 The third supplement trial of general fiber (resistant maltodextrin) reported no change in  
 292 *Bifidobacterium* spp. abundance within groups using FISH, although a significant increase  
 293 from baseline was reported for the intervention group on qPCR analysis (83). Finally, a food  
 294 study comparing intakes of wholegrains to refined grain products found no significant  
 295 difference in *Bifidobacterium* spp. abundance at the end of intervention period (78).  
 296 *Lactobacillus* spp. abundance was measured in 28 studies involving 867 participants. Data  
 297 from 24 studies (730 participants) was reported in a form suitable for meta-analysis (52, 55,  
 298 56, 60, 63-68, 73, 75, 76, 84, 87, 93, 96, 97, 99, 104, 105, 107, 111, 114). Dietary fiber led to a  
 299 significantly greater *Lactobacillus* spp. abundance compared with placebo/low fiber  
 300 comparators [SMD: 0.37 (95% CI: 0.07, 0.68),  $P = 0.02$ ]. However, heterogeneity was  
 301 considerable ( $I^2 = 80\%$ ), and was skewed by results from a single outlier study (66) [4.70 (95%  
 302 CI: 3.69, 5.70)]. A sensitivity analysis excluding this study produced a more homogenous  
 303 study population ( $I^2 = 49\%$ ), with a modest impact on the result [SMD: 0.22 (95% CI: 0.03,

0.41),  $P = 0.02$ ] (**Figure 3**). The outlier study (66) was excluded from subsequent subgroup analyses.

Subgroup analysis demonstrated interventions involving fiber supplements resulted in a significantly higher *Lactobacillus* spp. abundance compared with placebo/low fiber controls while substantially reducing study heterogeneity [SMD: 0.16 (95% CI: 0.01, 0.31),  $P = 0.04$ ,  $I^2 = 7\%$ ]. No significant differences in effect were found between food interventions and comparators [SMD: 0.35 (95% CI: -0.46, 1.16),  $P = 0.40$ ,  $I^2 = 84\%$ ].

Subgroup analysis of fiber types showed accepted prebiotic fiber interventions led to a significantly greater *Lactobacillus* spp. abundance compared with placebo/low fiber controls and further reduced heterogeneity [SMD: 0.34 (95% CI: 0.13, 0.55),  $P = 0.002$ ,  $I^2 = 0\%$ ] (**Figure 3**). There were no differences in effect in the candidate prebiotic [SMD: -0.06 (95% CI: -0.29, 0.16),  $P = 0.58$ ,  $I^2 = 0\%$ ] and general fiber [SMD: 0.22 (95% CI: -0.31, 0.75),  $P = 0.42$ ,  $I^2 = 74\%$ ] subgroups when compared with comparators.

Subgroup analysis of analysis method demonstrated dietary fiber led to significantly higher *Lactobacillus* spp. abundance compared with placebo/low fiber comparators when enumerated via culture [SMD: 0.61 (95% CI: 0.13, 1.08),  $P = 0.01$ ]. There were no significant differences between intervention and comparator when *Lactobacillus* spp. was detected using FISH, qPCR or sequencing (**Supplemental Table 6**). There were no differences in effect when sub-analyzing by intervention type or dose-response (**Supplemental Table 6**).

There were four studies that could not be pooled into the meta-analysis. A prebiotic supplementation trial of HMOs reported no difference in *Lactobacillus* spp. abundance between intervention and control groups (62). There was also no significant difference in *Lactobacillus* spp. reported in a wholegrain food intervention study compared with controls (78). Of the two remaining studies, there was higher *Lactobacillus* spp. abundance following xylo-oligosaccharide supplementation compared with placebo (69), and significant within-

group increases in *Lactobacillus* spp. abundance was demonstrated following manno-oligosaccharide supplementation (113).

Abundance of *F. prausnitzii* was measured in 15 studies investigating 566 participants. Thirteen studies (519 participants) were able to be meta-analyzed (53, 61, 67, 68, 74, 84, 88, 94, 99-101, 110, 112). There was no difference between dietary fiber compared with placebo/low fiber comparators for *F. prausnitzii* abundance [SMD: 0.14 (95% CI: -0.12, 0.39),  $P = 0.29$ ], with substantial heterogeneity between studies ( $I^2 = 68\%$ ) (**Figure 4**). Aside from trial design, no differences with respect to the pre-specified subgroups were found (**Supplemental Table 6**). Two studies reporting abundances of *F. prausnitzii* were unable to be pooled into the meta-analysis. Both studies measured the relative abundance of *F. prausnitzii* and reported only within-group changes, with one study reporting a decrease in abundance following supplementation of flaxseed mucilage (59), and the other reporting an increase in abundance following inulin supplementation (50).

Seven studies including 261 participants measured *Roseburia* spp. abundance. Four studies (189 participants) were included in the meta-analysis (49, 68, 79, 97). Dietary fiber had no effect on *Roseburia* spp. abundance compared with placebo/low fiber comparators [SMD: 0.33 (95% CI: -0.14, 0.80),  $P = 0.17$ ] although substantial heterogeneity was detected ( $I^2 = 70\%$ ) (**Figure 4**). Similar results were reported in the studies excluded from meta-analysis. No between or within-group differences were detected between intervention and placebo groups in two prebiotic fiber supplement trials (50, 62). A third trial found the relative abundance of *Roseburia* spp. was lower following inulin supplementation compared with control at end of intervention, although significance was not reported (115).

Two studies of 32 participants measured *E. hallii* abundance. These results could not be statistically pooled because one study did not report data in a suitable form. One study

reported no within-group difference in *E. hallii* abundance (50, 62), the other reported a significant decrease in *E. hallii* abundance compared with placebo (49). *E. rectale* was measured in three studies including 42 participants. Two studies (30 participants) were suitable for meta-analysis (84, 101). Dietary fiber did not impact on *E. rectale* abundance compared with placebo/low fiber comparators [SMD: -0.26 (95% CI: -1.20, 0.67),  $P = 0.58$ ] and substantial heterogeneity was detected ( $I^2 = 75\%$ ) (**Figure 4**). The study not eligible for meta-analysis was an inulin supplementation trial which reported no difference for within-group effects for *E. rectale* abundance (50). *R. bromii* abundance was measured in three studies encompassing 76 participants, of which all were suitable for meta-analysis (49, 81, 101). Dietary fiber had no effect on *R. bromii* abundance compared with placebo/low fiber comparators [SMD: 0.15 (95% CI: -0.15, 0.45),  $P = 0.33$ ], with no heterogeneity detected ( $I^2 = 0\%$ ) (**Figure 4**).

#### **Dietary fiber and short-chain fatty acids**

A total of 25 studies of 870 participants reported between-group differences in fecal SCFA concentration following fiber intervention (52, 53, 55, 59, 63, 64, 66-68, 71, 73, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112, 115). Fecal SCFA concentration was determined through gas-liquid chromatography in all but one study (90) where high-performance liquid chromatography was used.

Total fecal SCFA concentration was measured in 13 studies encompassing 406 participants (52, 55, 59, 63, 64, 67, 73, 80, 82, 84, 86, 91, 94). Dietary fiber had no effect on total SCFA concentration compared with placebo/low fiber comparators [SMD: 0.11 (95% CI: -0.05, 0.27),  $P = 0.19$ ], with similar intervention effects across studies ( $I^2 = 0\%$ ).

Fecal acetate concentration was reported in 18 studies involving 657 participants (52, 53, 63, 66, 71, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112). There was no difference in fecal

377 acetate following fiber intervention compared with placebo/low fiber comparators [SMD: 0.28  
 378 (95% CI: -0.08, 0.63),  $P = 0.13$ ] with substantial heterogeneity between studies ( $I^2 = 86$ ).  
 379 The effect of fiber intervention on fecal propionate concentration was reported in 19 studies of  
 380 677 participants (52, 53, 63, 66, 71, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112, 115).  
 381 No differences were found between fecal propionate and comparators [SMD: -0.01 (95% CI: -  
 382 0.20, 0.22),  $P = 0.95$ ], with moderate heterogeneity detected ( $I^2 = 61\%$ ).  
 383 The effect of fiber intervention on fecal butyrate concentration was reported in 20 studies of  
 384 712 participants (52, 53, 59, 63, 66, 71, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112,  
 385 115). Fecal butyrate was significantly higher following fiber intervention compared with  
 386 placebo/low fiber comparators [SMD: 0.24 (95% CI: 0.00, 0.47),  $P = 0.05$ ], although  
 387 considerable heterogeneity was present ( $I^2 = 70\%$ ).  
 388 Of the studies evaluating differences in fecal SCFA concentration following fiber intervention  
 389 compared with placebo/low fiber comparators, 13 studies expressed mean SCFA  
 390 concentrations per wet weight of feces (52, 53, 66, 67, 71, 73, 74, 77, 82, 90, 91, 96, 115), 10  
 391 studies as dry weight of feces (55, 59, 63, 64, 68, 80, 93, 94, 103, 112), one study as molar  
 392 ratio (84), and one study as a combination of wet weight of feces and molar ratio (86).  
 393 Additional subgroup analyses were performed to compare differences in fecal SCFA  
 394 concentrations when expressed as wet weight compared with dry weight (**Supplemental**  
 395 **Table 7**). Fiber intervention led to significantly higher fecal concentrations of total SCFA,  
 396 acetate and butyrate compared with comparators when expressed per wet weight of feces.  
 397 However, there were no significant differences when mean SCFA concentrations were  
 398 expressed per dry weight of feces. Study heterogeneity was considerably greater for fecal  
 399 acetate and butyrate, but not total fecal SCFA concentrations when expressed as wet compared  
 400 with dry wet of feces. There were no differences in effect based on analysis method for fecal

propionate concentrations, although heterogeneity was greater when results were expressed per wet weight of feces (**Supplemental Table 7**).

### **Differences in intervention effects based on trial design**

There were differences in intervention effects in subgroup analyses depending upon trial design. Dietary fiber led to significantly lower  $\alpha$ -diversity compared with placebo/low fiber comparators in crossover design trials, where  $\alpha$ -diversity was reported using Shannon diversity index [MD: -0.10 (95% CI: -0.19, -0.01),  $P = 0.03$ ], while there was no difference in  $\alpha$ -diversity in parallel design trials [MD: -0.03 (95% CI: -0.57, 0.51),  $P = 0.91$ ] (**Supplemental Table 6**). The presence and duration of washout periods were inconsistent across the three crossover trials included this analysis. One study did not include a wash out period (84), and wash out periods lasted 14 (75) and 21 days (88) in the other two. Regarding bacterial abundances however, intervention effects were significant in parallel trials but not in crossover trials for *Lactobacillus* and *Roseburia* spp. and *F. prausnitzii*, but not for *Bifidobacterium* spp. (**Supplemental Table 6**). Statistical heterogeneity was lower in crossover trials compared with parallel trials for  $\alpha$ -diversity reported using Shannon diversity index, *Bifidobacterium* and *Lactobacillus* spp., as well as *F. prausnitzii*, but there was no difference in statistical heterogeneity for *Roseburia* spp. (**Supplemental Table 6**).

### **Risk of bias**

The risk of bias was low-to-moderate across the 64 included studies (**Supplemental Figure 1**). Selection bias was unclear in most studies. Random sequence generation and allocation concealment were adequately described by 26% (59-62, 70-72, 77, 79, 80, 84, 86, 94, 103, 113-115) and 16% (59, 61, 62, 70, 77, 79, 80, 86, 94, 115) of studies, respectively. There was low risk of bias across included studies regarding performance and detection bias, as most trials investigated objective outcomes and incorporated a double-blind design. Attrition bias was adequately addressed by only 41% (54-58, 62, 67, 69, 71, 74-76, 79, 82, 86-89, 92, 93, 98,

99, 105, 107, 108, 110) of the included studies. Selective reporting was unclear in the majority of studies. Published protocols or clinical registrations were reported by only 26% (59, 61, 68-70, 75, 77-80, 86, 97, 100-102, 110, 115) of included studies. Bias related to control of dietary intake was unclear in half of included studies (55%) (54, 56-60, 62, 64-67, 71, 72, 74, 78, 80, 81, 83, 85-93, 96, 98, 102, 103, 105, 108, 110, 115), while even fewer studies were judged to have a low risk of bias regarding dietary advice and assessment of dietary compliance (33%) (52, 55, 63, 68, 69, 73, 75, 76, 79, 82, 84, 94, 97, 99, 104, 106, 107, 111-114). Furthermore, 13% (53, 61, 70, 77, 95, 100, 101, 109) of studies did not provide dietary advice or assess intake, and were judged to have a high risk of bias relating to the potential influence of background dietary intake.

#### **Reporting bias**

Funnel plots were generated for abundances of *Bifidobacterium* spp.; *Lactobacillus* spp.; *F. prausnitzii*; and total SCFA; acetate; propionate; and butyrate concentrations. Visual inspection found no evidence of funnel plot asymmetry, indicating reporting bias was unlikely (Supplemental Figures 2-7).

## DISCUSSION

This systematic review and meta-analysis found dietary fiber intervention had no effect on the diversity of the gut microbiota but did increase abundance of *Bifidobacterium* and *Lactobacillus* spp. as well as fecal butyrate concentration in healthy adults.

The lack of effect on  $\alpha$ -diversity of the gut microbiota found in this review is similar to other dietary interventions documented in the literature. For instance, controlled feeding studies lasting four days to three weeks found that despite significant changes to fiber intake, there was no effect on microbial diversity (35-37). These findings suggest that short-term dietary interventions are unlikely to facilitate changes in the  $\alpha$ -diversity of the gut microbiota. Indeed, study design is likely important, as subgroup analysis demonstrated different effects between crossover and parallel trials. The lower  $\alpha$ -diversity between fiber and control groups in crossover trials may be related to a lack of or insufficient wash-out between interventions, as well as potential differences in the microbiota and habitual diet of individuals at baseline. These null findings are in contrast to the findings from observational studies that report a correlation between fiber intakes in habitual diet and diversity of the gut microbiota, for example in studies comparing agrarian dietary habits with Western populations (38, 39). Interestingly, a positive correlation has also been reported between dietary diversity and microbiota diversity (116). Taken together, long term dietary diversity as opposed to changes in isolated nutrients or foods over a short period of time may be a stronger driver of microbial diversity. It must also be noted that the stability of the gut microbiota, as well as the abundances and metabolites of the individual members of the microbial community, also contribute to maintaining an ecosystem that promotes health (117, 118). Therefore, the totality of findings here, including that microbial diversity was not compromised, support the favorable effects of dietary fiber on the gut microbiota.



In regard to particular bacterial groups, this review demonstrated dietary fiber interventions involving accepted prebiotic fibers led to higher abundance of *Bifidobacterium* and *Lactobacillus* species. These results support the selectivity criteria of the prebiotic concept, where the host microorganisms selectively utilize the prebiotic fibers as substrates, which may confer health benefits to the host (32). However, candidate prebiotic interventions produced different effects on the abundance of these two genera, with significant effects demonstrated for *Bifidobacterium* but not *Lactobacillus* species. This may represent differences in substrate preferences between the two genera, where *Bifidobacterium* spp. may be less discriminating than *Lactobacillus* spp. regarding fermentation substrates (119, 120). Conversely, fibers not classified as accepted or candidate prebiotics, here termed general fibers, did not impact the abundance of these taxa. This may be due to the heterogeneity of the general fibers, including their degree of polymerization, viscosity and fermentability, whereas accepted and candidate prebiotic fibers are mostly highly fermentable oligosaccharides (29, 30).

Subgroup analysis separating the effect of food vs supplement interventions showed food interventions had no effect on *Bifidobacterium* and *Lactobacillus* species. This result may be attributed to a lack of statistical power, due to the food interventions comprising a relatively small number of low sample size studies (10 studies, 301 participants; 4 studies, 127 participants). It must also be noted that most of the trials employing food interventions supplemented with grain and cereal foods to increase fiber intake (71, 78, 79, 82, 85, 89, 96, 98, 104). Therefore, the food interventions evaluated may be more representative of grains and cereals *per se* rather than a diverse range of fibrous foods.

Interestingly, there were no differences in the effect of dietary fiber interventions on *Bifidobacterium* spp. abundance with varying doses of fiber. Dietary fiber intervention led to an effect at all levels of consumption in subgroup analysis ( $\leq 5\text{g}$ , 5-10g,  $>10\text{g}$ ) with no discernible gradient in effectiveness, suggesting fewer than 5 grams of dietary fiber is

sufficient. This may represent a potential limit to the amount of fiber that can be fermented by *Bifidobacterium* species. The lack of a dose-response effect may also be attributed to the percentage increase in fiber intake from baseline rather than the intervention dose, which was unable to be accounted for in this review due to the inconsistent reporting of baseline values across included studies. This requires further clarification but lower dose supplementation may be advantageous in patients who experience GI symptoms with higher fiber loads.

There was more variability in intervention effects for abundances of *Bifidobacterium* spp. ( $I^2 = 85\%$ ) compared with *Lactobacillus* spp. ( $I^2 = 49\%$ ). While this may be related to differences in the accuracy of techniques used to determine specific bacterial abundances (121, 122), there were no differences in effect based on analysis method for *Bifidobacterium* species. Another plausible explanation is the differences in nutrient requirements of these taxa as discussed previously. Furthermore, ‘responder and non-responder’ effects for *Bifidobacterium* spp. abundance, which have been shown previously (97, 123, 124), may be impacted by individual host factors, such as differences in baseline abundances (124), or the presence/absence of specific strains of *Bifidobacterium* able to utilize the particular fiber under investigation.

There were differences in intervention effects based on trial design, with parallel design studies demonstrating stronger intervention effects and greater statistical heterogeneity compared with crossover design studies for several outcomes. This may in part be due to inter-individual differences in microbiota composition as well as carry-over effects from a lack of or insufficient wash-out periods in the crossover studies as discussed previously.

There was no effect of dietary fiber interventions on abundance of other commonly measured bacterial groups (e.g. *F. prausnitzii*), suggesting these species may be stimulated by dietary components other than fiber, such as polyols and polyphenols (125). However, the number of studies evaluating species of other bacterial groups was small, and therefore further studies are needed to investigate the effect of fiber and other dietary components on these groups.

The higher fecal concentration of butyrate following fiber intervention highlights the ability of dietary fiber to beneficially modulate the metabolic outputs of the gut microbiota. This is likely due to cross-feeding interactions between butyrate producers with *Bifidobacterium* and *Lactobacillus* species, which are noted lactate and acetate producers (25, 120, 126). As the preferred energy source for colonic epithelial cells, butyrate is a microbial by-product that is of particular interest to host health, exhibiting a wide spectrum of positive effects, such as inhibiting colonic carcinogenesis and ameliorating mucosal inflammation (31, 127, 128). However, it is acknowledged that the variability in the reporting of SCFA results may limit the applicability of these findings, particularly when considering the variance in results when expressed as wet compared with dry weight of feces.

This study is the first systematic review and meta-analysis to assess the effect of dietary fiber intervention on gut microbiota composition. Major strengths of this study include its robust design, comprehensive search strategies, and the use of two independent reviewers.

It is acknowledged this study has some limitations. Firstly, there were only a limited number of studies reporting the primary outcome of  $\alpha$ -diversity, and a small proportion presenting data using the same diversity indices. Secondly, baseline fiber intake was not able to be accounted for due to the paucity of reporting by included studies. Furthermore, included studies sampled feces as a surrogate for gut microbiota profile, and although feces are a common sampling route, the microbial composition of feces differs from the mucosal microbiota (10, 11), which is in closer contact with the host and may be more important when considering the relationship between microbiota and disease pathophysiology or outcomes. Finally, the limited number of taxa assessed in the review may not convey the overall effect elicited by dietary fiber intervention on gut microbiota composition and metabolic outputs, although the selection of taxa was guided by the available literature. Thus, the taxa selected may be more representative of the scope of research in the field to date, rather than a limitation of the review.

540 Dietary fiber intervention leads to a higher abundance of fecal *Bifidobacterium* and  
541 *Lactobacillus* spp., as well as higher fecal concentration of butyrate compared with  
542 placebo/low fiber comparators. Accepted prebiotic fibers had an effect on the abundances of  
543 both *Bifidobacterium* and *Lactobacillus* spp. while candidate prebiotic fibers had an effect on  
544 *Bifidobacterium* spp. abundance but not *Lactobacillus* species. General fibers appear to have a  
545 limited effect on gut microbiota composition. Although the diversity of the gut microbiota,  
546 abundances of other commonly measured bacterial groups and concentration of other fecal  
547 SCFAs were not significantly different compared with controls following dietary fiber  
548 intervention, it is worth noting that a short-term increase in fiber intake does not appear to be  
549 rate-limiting to these outcomes. These results further support the favorable effects of dietary  
550 fiber and contribute to our understanding of its effect on the gut microbiota.

551 Future RCTs investigating the effect of fiber on the gut microbiota should adjust for  
552 participants' baseline microbiota composition and dietary characteristics as well as controlling  
553 for dietary intake in order to determine the precise effect of dietary fiber. Scope may also need  
554 to be broadened to evaluate taxa than that considered here, including the eukaryote (e.g. fungi)  
555 members of the gut microbiota. Additionally, longer duration studies are needed to better  
556 assess the chronic effect of fiber on microbiota diversity.

557 **Author contributions**

558 The author's responsibilities were as follows – HS and KC: initiated the study; DS, KW, HS,  
559 MR, KW and KC: developed the protocol; DS and HS: performed eligibility screening and  
560 data extraction; DS and JK: analyzed the data and performed the statistical analysis; DS KW,  
561 MR, MM, JK, ES, HS and KC: interpreted the data; DS: wrote the initial manuscript; and KW,  
562 MR, MM, GH, JK, ES, HS and KC: critically revised the manuscript. All authors read and  
563 approved the final manuscript.

564 **Competing interests**

565 None declared.

566 **Acknowledgements**

567 The authors wish to thank David Honeyman for assisting with the development of the search  
568 strategy. Many thanks to the authors of included studies who provided outcome data necessary  
569 for the extraction of data of the variables included in the meta-analyses.

## REFERENCES

1. Sender R, Fuchs S, Milo R. Revised Estimates for the Number of Human and Bacteria Cells in the Body. *PLoS biology* 2016;14(8):e1002533. doi: 10.1371/journal.pbio.1002533.
2. Marchesi JR, Adams DH, Fava F, Hermes GDA, Hirschfield GM, Hold G, Quraishi MN, Kinross J, Smidt H, Tuohy KM, et al. The gut microbiota and host health: a new clinical frontier. *Gut* 2016;65(2):330-9. doi: 10.1136/gutjnl-2015-309990.
3. Scott KP, Duncan SH, Flint HJ. Dietary fibre and the gut microbiota. *Nutrition Bulletin* 2008;33(3):201-11. doi: 10.1111/j.1467-3010.2008.00706.x.
4. LeBlanc JG, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Current opinion in biotechnology* 2013;24(2):160-8. doi: 10.1016/j.copbio.2012.08.005.
5. Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. *Nature reviews Immunology* 2009;9(5):313-23. doi: 10.1038/nri2515.
6. Sekirov I, Russell SL, Antunes LC, Finlay BB. Gut microbiota in health and disease. *Physiological reviews* 2010;90(3):859-904. doi: 10.1152/physrev.00045.2009.
7. Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell* 2012;148(6):1258-70. doi: 10.1016/j.cell.2012.01.035.
8. de Vos WM, de Vos EA. Role of the intestinal microbiome in health and disease: from correlation to causation. *Nutrition reviews* 2012;70 Suppl 1:S45-56. doi: 10.1111/j.1753-4887.2012.00505.x.
9. Zhernakova A, Kurilshikov A, Bonder MJ, Tigchelaar EF, Schirmer M, Vatanen T, Mujagic Z, Vila AV, Falony G, Vieira-Silva S, et al. Population-based metagenomics

- analysis reveals markers for gut microbiome composition and diversity. *Science* (New York, NY) 2016;352(6285):565-9. doi: 10.1126/science.aad3369.
10. Fraher MH, O'Toole PW, Quigley EM. Techniques used to characterize the gut microbiota: a guide for the clinician. *Nature reviews Gastroenterology & hepatology* 2012;9(6):312-22. doi: 10.1038/nrgastro.2012.44.
  11. Zoetendal EG, von Wright A, Vilpponen-Salmela T, Ben-Amor K, Akkermans AD, de Vos WM. Mucosa-associated bacteria in the human gastrointestinal tract are uniformly distributed along the colon and differ from the community recovered from feces. *Applied and environmental microbiology* 2002;68(7):3401-7.
  12. Lozupone CA, Knight R. Species divergence and the measurement of microbial diversity. *FEMS microbiology reviews* 2008;32(4):557-78. doi: 10.1111/j.1574-6976.2008.00111.x.
  13. Morgan XC, Huttenhower C. Chapter 12: Human microbiome analysis. *PLoS computational biology* 2012;8(12):e1002808. doi: 10.1371/journal.pcbi.1002808.
  14. Tojo R, Suarez A, Clemente MG, de los Reyes-Gavilan CG, Margolles A, Gueimonde M, Ruas-Madiedo P. Intestinal microbiota in health and disease: role of bifidobacteria in gut homeostasis. *World journal of gastroenterology* 2014;20(41):15163-76. doi: 10.3748/wjg.v20.i41.15163.
  15. Bottacini F, Ventura M, van Sinderen D, O'Connell Motherway M. Diversity, ecology and intestinal function of bifidobacteria. *Microbial cell factories* 2014;13 Suppl 1:S4. doi: 10.1186/1475-2859-13-s1-s4.
  16. Rossi M, Amaretti A, Raimondi S. Folate production by probiotic bacteria. *Nutrients* 2011;3(1):118-34. doi: 10.3390/nu3010118.
  17. Satokari RM, Vaughan EE, Smidt H, Saarela M, Matto J, de Vos WM. Molecular approaches for the detection and identification of bifidobacteria and lactobacilli in the

- human gastrointestinal tract. *Systematic and applied microbiology* 2003;26(4):572-84. doi: 10.1078/072320203770865882.
18. Wells JM. Immunomodulatory mechanisms of lactobacilli. *Microbial cell factories* 2011;10 Suppl 1:S17. doi: 10.1186/1475-2859-10-s1-s17.
  19. Flint HJ, Duncan SH, Louis P. The impact of nutrition on intestinal bacterial communities. *Current opinion in microbiology* 2017;38:59-65. doi: 10.1016/j.mib.2017.04.005.
  20. Flint HJ, Duncan SH, Scott KP, Louis P. Links between diet, gut microbiota composition and gut metabolism. *The Proceedings of the Nutrition Society* 2015;74(1):13-22. doi: 10.1017/s0029665114001463.
  21. Ze X, Duncan SH, Louis P, Flint HJ. *Ruminococcus bromii* is a keystone species for the degradation of resistant starch in the human colon. *The ISME journal* 2012;6(8):1535-43. doi: 10.1038/ismej.2012.4.
  22. Ze X, Le Mougen F, Duncan SH, Louis P, Flint HJ. Some are more equal than others: the role of "keystone" species in the degradation of recalcitrant substrates. *Gut microbes* 2013;4(3):236-40. doi: 10.4161/gmic.23998.
  23. Scott KP, Antoine J-M, Midtvedt T, van Hemert S. Manipulating the gut microbiota to maintain health and treat disease. 2015 2015;26. doi: 10.3402/mehd.v26.25877.
  24. O'Keefe SJ. Diet, microorganisms and their metabolites, and colon cancer. *Nature reviews Gastroenterology & hepatology* 2016;13(12):691-706. doi: 10.1038/nrgastro.2016.165.
  25. Rivière A, Selak M, Lantin D, Leroy F, De Vuyst L. Bifidobacteria and Butyrate-Producing Colon Bacteria: Importance and Strategies for Their Stimulation in the Human Gut. *Frontiers in Microbiology* 2016;7:979. doi: 10.3389/fmicb.2016.00979.



26. FAO/WHO. CODEX Alimentarius (CODEX) Guidelines on Nutrition Labeling CAC/GL 2–1985. 2013.
27. Federalregister.gov [Internet]. Washington (DC): Food and Drug Administration Health and Human Services. Federal Register. Food labeling: revision of the nutrition and supplement facts labels (21 CFR 101) [Internet]. c. May 2016 [cited 2018 Jan 10]. Available from: <https://www.federalregister.gov/documents/2016/05/27/2016-11867/food-labelingrevision-of-the-nutrition-and-supplement-facts-labels>,.
28. Eswaran S, Muir J, Chey WD. Fiber and functional gastrointestinal disorders. The American journal of gastroenterology 2013;108(5):718-27. doi: 10.1038/ajg.2013.63.
29. Holscher HD. Dietary fiber and prebiotics and the gastrointestinal microbiota. Gut microbes 2017;8(2):172-84. doi: 10.1080/19490976.2017.1290756.
30. McRorie JW, Jr., McKeown NM. Understanding the Physics of Functional Fibers in the Gastrointestinal Tract: An Evidence-Based Approach to Resolving Enduring Misconceptions about Insoluble and Soluble Fiber. Journal of the Academy of Nutrition and Dietetics 2017;117(2):251-64. doi: 10.1016/j.jand.2016.09.021.
31. Wong JM, de Souza R, Kendall CW, Emam A, Jenkins DJ. Colonic health: fermentation and short chain fatty acids. Journal of clinical gastroenterology 2006;40(3):235-43.
32. Gibson GR, Hutkins R, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, Scott K, Stanton C, Swanson KS, Cani PD, et al. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. Nature reviews Gastroenterology & hepatology 2017;advance online publication. doi: 10.1038/nrgastro.2017.75.
33. Gibson GR, Scott KP, Rastall RA, Tuohy KM, Hotchkiss A, Dubert-Ferrandon A, Gareau M, Murphy EF, Saulnier D, Loh G, et al. Dietary prebiotics: current status and

- new definition. *Food Science & Technology Bulletin: Functional Foods* 2010;7(1):1-19. doi: 10.1616/1476-2137.15880.
34. Roberfroid M, Gibson GR, Hoyles L, McCartney AL, Rastall R, Rowland I, Wolvers D, Watzl B, Szajewska H, Stahl B, et al. Prebiotic effects: metabolic and health benefits. *Br J Nutr* 2010;104 Suppl 2:S1-63. doi: 10.1017/s0007114510003363.
  35. David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, Ling AV, Devlin AS, Varma Y, Fischbach MA, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014;505(7484):559-63. doi: 10.1038/nature12820.
  36. Walker AW, Ince J, Duncan SH, Webster LM, Holtrop G, Ze X, Brown D, Stares MD, Scott P, Bergerat A, et al. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *The ISME journal* 2011;5(2):220-30. doi: 10.1038/ismej.2010.118.
  37. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen Y-Y, Keilbaugh SA, Bewtra M, Knights D, Walters WA, Knight R, et al. Linking Long-Term Dietary Patterns with Gut Microbial Enterotypes. *Science (New York, NY)* 2011;334(6052):105-8. doi: 10.1126/science.1208344.
  38. De Filippo C, Cavalieri D, Di Paola M, Ramazzotti M, Poullet JB, Massart S, Collini S, Pieraccini G, Lionetti P. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proceedings of the National Academy of Sciences of the United States of America* 2010;107(33):14691-6. doi: 10.1073/pnas.1005963107.
  39. Schnorr SL, Candela M, Rampelli S, Centanni M, Consolandi C, Basaglia G, Turrioni S, Biagi E, Peano C, Severgnini M, et al. Gut microbiome of the Hadza hunter-gatherers. *Nature Communications* 2014;5:3654. doi: 10.1038/ncomms4654

<http://www.nature.com/articles/ncomms4654#supplementary-information>.

40. Falony G, Joossens M, Vieira-Silva S, Wang J, Darzi Y, Faust K, Kurilshikov A, Bonder MJ, Valles-Colomer M, Vandeputte D, et al. Population-level analysis of gut microbiome variation. *Science* (New York, NY) 2016;352(6285):560-4. doi: 10.1126/science.aad3503.
41. Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P, Stewart LA. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Systematic Reviews* 2015;4(1):1-9. doi: 10.1186/2046-4053-4-1.
42. Higgins JPT, Green S. *Cochrane Handbook for Systematic Reviews of Interventions*: Wiley, 2011.
43. Elmagarmid A, Fedorowicz Z, Hammady H, Ilyas I, Khabsa M, Ouzzani M. Rayyan: a systematic reviews web app for exploring and filtering searches for eligible studies for Cochrane Reviews. *Evidence-Informed Public Health: Opportunities and Challenges Abstracts of the 22nd Cochrane Colloquium*. Hyderabad, India: John Wiley & Sons, 2014.
44. Higgins JPT, Deeks JJ, Altman DG. *Special Topics in Statistics*. Edition ed. *Cochrane Handbook for Systematic Reviews of Interventions*: John Wiley & Sons, Ltd, 2008:481-529.
45. Higgins JPT, Altman DG. *Assessing Risk of Bias in Included Studies*. Edition ed. *Cochrane Handbook for Systematic Reviews of Interventions*: John Wiley & Sons, Ltd, 2008:187-241.
46. Higgins JPT, Deeks JJ. *Selecting Studies and Collecting Data*. Edition ed. *Cochrane Handbook for Systematic Reviews of Interventions*: John Wiley & Sons, Ltd, 2008:151-85.

47. Elbourne DR, Altman DG, Higgins JP, Curtin F, Worthington HV, Vail A. Meta-analyses involving cross-over trials: methodological issues. *International journal of epidemiology* 2002;31(1):140-9.
48. Sterne JAC, Egger M, Moher D. Addressing Reporting Biases. Edition ed. *Cochrane Handbook for Systematic Reviews of Interventions*: John Wiley & Sons, Ltd, 2008:297-333.
49. Hooda S, Vester Boler BM, Rossoni Sero MC, Brulc JM, Staeger MA, Boileau TW, Dowd SE, Fahey Jr GC, Swanson KS. 454 pyrosequencing reveals a shift in fecal microbiota of healthy adult men consuming polydextrose or soluble corn fiber. *Journal of Nutrition* 2012;142(7):1259-65.
50. Ramirez-Farias C, Slezak K, Fuller Z, Duncan A, Holtrop G, Louis P. Effect of inulin on the human gut microbiota: Stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*. *British Journal of Nutrition* 2009;101(4):541-50.
51. Vanegas SM, Meydani M, Barnett JB, Goldin B, Kane A, Rasmussen H, Brown C, Vangay P, Knights D, Jonnalagadda S, et al. Substituting whole grains for refined grains in a 6-wk randomized trial has a modest effect on gut microbiota and immune and inflammatory markers of healthy adults. *American Journal of Clinical* 2017;105(3):635-50.
52. Alles MS, Hartemink R, Meyboom S, Harryvan JL, Van Laere KMJ, Nagengast FM, Hautvast JGAJ. Effect of transgalactooligosaccharides on the composition of the human intestinal microflora and on putative risk markers for colon cancer. *American Journal of Clinical Nutrition* 1999;69(5):980-91.
53. Beards E, Tuohy K, Gibson G. A human volunteer study to assess the impact of confectionery sweeteners on the gut microbiota composition. *British Journal of Nutrition* 2010;104(5):701-8.

54. Bouhnik Y, Flourié B, Riottot M, Bisetti N, Gailing MF, Guibert A, Bornet F, Rambaud JC. Effects of fructo-oligosaccharides ingestion on fecal bifidobacteria and selected metabolic indexes of colon carcinogenesis in healthy humans. *Nutrition and Cancer* 1996;26(1):21-9.
55. Bouhnik Y, Raskine L, Champion K, Andrieux C, Penven S, Jacobs H, Simoneau G. Prolonged administration of low-dose inulin stimulates the growth of bifidobacteria in humans. *Nutrition Research* 2007;27(4):187-93.
56. Bouhnik Y, Raskine L, Simoneau G, Paineau D, Bornet F. The capacity of short-chain fructo-oligosaccharides to stimulate faecal bifidobacteria: A dose-response relationship study in healthy humans. *Nutrition journal* 2006;5.
57. Bouhnik Y, Raskine L, Simoneau G, Vicaud E, Neut C, Flourié B, Brouns F, Bornet FR. The capacity of nondigestible carbohydrates to stimulate fecal bifidobacteria in healthy humans: A double-blind, randomized, placebo-controlled, parallel-group, dose-response relation study. *American Journal of Clinical Nutrition* 2004;80(6):1658-64.
58. Bouhnik Y, Vahedi K, Achour L, Attar A, Salfati J, Pochart P, Marteau P, Flourié B, Bornet F, Rambaud JC. Short-chain fructo-oligosaccharide administration dose-dependently increases fecal bifidobacteria in healthy humans. *Journal of Nutrition* 1999;129(1):113-6.
59. Brahe L, Chatelier E, Prifti E, Pons N, Kennedy S, Blædel T, Håkansson J, Dalsgaard T, Hansen T, Pedersen O, et al. Dietary modulation of the gut microbiota--a randomised controlled trial in obese postmenopausal women. *The British journal of nutrition* 2015;114(3):406-17.
60. Calame W, Weseler AR, Viebke C, Flynn C, Siemensma AD. Gum arabic establishes prebiotic functionality in healthy human volunteers in a dose-dependent manner. *British Journal of Nutrition* 2008;100(6):1269-75.

61. Dewulf EM, Cani PD, Claus SP, Fuentes S, Puylaert PGB, Neyrinck AM, Bindels LB, De Vos WM, Gibson GR, Thissen JP, et al. Insight into the prebiotic concept: Lessons from an exploratory, double blind intervention study with inulin-type fructans in obese women. *Gut* 2013;62(8):1112-21.
62. Elison E, Vigsnaes LK, Rindom Krogsgaard L, Rasmussen J, Sørensen N, McConnell B, Hennet T, Sommer MOA, Bytzer P. Oral supplementation of healthy adults with 2'-O-fucosyllactose and lacto-N-neotetraose is well tolerated and shifts the intestinal microbiota. *British Journal of Nutrition* 2016;0:1-13.
63. Fastinger ND, Karr-Lilienthal LK, Spears JK, Swanson KS, Zinn KE, Nava GM, Ohkuma K, Kanahori S, Gordon DT, Fahey Jr GC. A novel resistant maltodextrin alters gastrointestinal tolerance factors, fecal characteristics, and fecal microbiota in healthy adult humans. *Journal of the American College of Nutrition* 2008;27(2):356-66.
64. Finegold S, Li Z, Summanen P, Downes J, Thames G, Corbett K, Dowd S, Krak M, Heber D. Xylooligosaccharide increases bifidobacteria but not lactobacilli in human gut microbiota. *Food & function* 2014;5(3):436-45.
65. Gopal PK, Prasad J, Gill HS. Effects of the consumption of *Bifidobacterium lactis* HN019 (DR10™) and galacto-oligosaccharides on the microflora of the gastrointestinal tract in human subjects. *Nutrition Research* 2003;23(10):1313-28.
66. Jie Z, Bang-Yao L, Ming-Jie X, Hai-Wei L, Zu-Kang Z, Ting-Song W, Craig S. Studies on the effects of polydextrose intake on physiologic functions in Chinese people. *The American journal of clinical nutrition* 2000;72(6):1503-9.
67. Kleessen B, Schwarz S, Boehm A, Fuhrmann H, Richter A, Henle T, Krueger M. Jerusalem artichoke and chicory inulin in bakery products affect faecal microbiota of healthy volunteers. *British Journal of Nutrition* 2007;98(3):540-9.

68. Lecerf JM, Dépeint F, Clerc E, Dugenet Y, Niamba CN, Rhazi L, Cayzeele A, Abdelnour G, Jaruga A, Younes H, et al. Xylo-oligosaccharide (XOS) in combination with inulin modulates both the intestinal environment and immune status in healthy subjects, while XOS alone only shows prebiotic properties. *British Journal of Nutrition* 2012;108(10):1847-58.
69. Lin SH, Chou LM, Chien YW, Chang JS, Lin CI. Prebiotic Effects of Xylooligosaccharides on the Improvement of Microbiota Balance in Human Subjects. *Gastroenterology Research and Practice* 2016;2016.
70. Lomax AR, Cheung LVY, Tuohy KM, Noakes PS, Miles EA, Calder PC.  $\beta$ 2-1 Fructans have a bifidogenic effect in healthy middle-aged human subjects but do not alter immune responses examined in the absence of an in vivo immune challenge: Results from a randomised controlled trial. *British Journal of Nutrition* 2012;108(10):1818-28.
71. Nemoto H, Ikata K, Arimochi H, Iwasaki T, Ohnishi Y, Kuwahara T, Kataoka K. Effects of fermented brown rice on the intestinal environments in healthy adult. *Journal of Medical Investigation* 2011;58(3):235-45.
72. Pallav K, Dowd SE, Villafuerte J, Yang X, Kabbani T, Hansen J, Dennis M, Leffler DA, Kelly CP. Effects of polysaccharopeptide from *Trametes versicolor* and amoxicillin on the gut microbiome of healthy volunteers: A randomized clinical trial. *Gut microbes* 2014;5(4).
73. Pasman W, Wils D, Saniez MH, Kardinaal A. Long-term gastrointestinal tolerance of NUTRIOSE®FB in healthy men. *European journal of clinical nutrition* 2006;60(8):1024-34.

74. Ramnani P, Gaudier E, Bingham M, Van Bruggen P, Tuohy KM, Gibson GR. Prebiotic effect of fruit and vegetable shots containing Jerusalem artichoke inulin: A human intervention study. *British Journal of Nutrition* 2010;104(2):233-40.
75. Tap J, Furet JP, Bensaada M, Philippe C, Roth H, Rabot S, Lakhdari O, Lombard V, Henrissat B, Corthier G, et al. Gut microbiota richness promotes its stability upon increased dietary fibre intake in healthy adults. *Environmental microbiology* 2015;17(12):4954-64.
76. Wu WT, Cheng HC, Chen HL. Ameliorative effects of konjac glucomannan on human faecal beta-glucuronidase activity, secondary bile acid levels and faecal water toxicity towards Caco-2 cells. *British Journal of Nutrition* 2011;105(4):593-600.
77. Alfa MJ, Strang D, Tappia PS, Graham M, Van Domselaar G, Forbes JD, Laminman V, Olson N, DeGagne P, Bray D, et al. A randomized trial to determine the impact of a digestion resistant starch composition on the gut microbiome in older and mid-age adults. *Clinical* 2017.
78. Cooper DN, Kable ME, Marco ML, De Leon A, Rust B, Baker JE, Horn W, Burnett D, Keim NL. The Effects of Moderate Whole Grain Consumption on Fasting Glucose and Lipids, Gastrointestinal Symptoms, and Microbiota. *Nutrients* 2017;9(2). doi: 10.3390/nu9020173.
79. Karl JP, Meydani M, Barnett JB, Vanegas SM, Goldin B, Kane A, Rasmussen H, Saltzman E, Vangay P, Knights D, et al. Substituting whole grains for refined grains in a 6-wk randomized trial favorably affects energy-balance metrics in healthy men and postmenopausal women. *American Journal of Clinical Nutrition* 2017;105(3):589-99.
80. Salden BN, Troost FJ, Wilms E, Truchado P, Vilchez-Vargas R, Pieper DH, Jáuregui R, Marzorati M, van de Wiele T, Possemiers S, et al. Reinforcement of intestinal



epithelial barrier by arabinoxylans in overweight and obese subjects: A randomized controlled trial. Arabinoxylans in gut barrier. Clinical 2017.

81. Abell GCJ, Cooke CM, Bennett CN, Conlon MA, McOrist AL. Phylotypes related to *Ruminococcus bromii* are abundant in the large bowel of humans and increase in response to a diet high in resistant starch. *FEMS Microbiology Ecology* 2008;66(3):505-15.
82. Ampatzoglou A, Atwal KK, Maidens CM, Williams CL, Ross AB, Thielecke F, Jonnalagadda SS, Kennedy OB, Yaqoob P. Increased whole grain consumption does not affect blood biochemistry, body composition, or gut microbiology in healthy, low-habitual whole grain consumers. *Journal of Nutrition* 2015;145(2):215-21.
83. Baer DJ, Stote KS, Henderson T, Paul DR, Okuma K, Tagami H, Kanahori S, Gordon DT, Rumpler WV, Ukhanova M, et al. The metabolizable energy of dietary resistant maltodextrin is variable and alters fecal microbiota composition in adult men. *Journal of Nutrition* 2014;144(7):1023-9.
84. Boler B, Seroo M, Bauer L, Staeger M, Boileau T, Swanson K, Fahey G. Digestive physiological outcomes related to polydextrose and soluble maize fibre consumption by healthy adult men. *The British journal of nutrition* 2011;106(12):1864-71.
85. Carvalho-Wells AL, Helmolz K, Nodet C, Molzer C, Leonard C, McKevith B, Thielecke F, Jackson KG, Tuohy KM. Determination of the in vivo prebiotic potential of a maize-based whole grain breakfast cereal: A human feeding study. *British Journal of Nutrition* 2010;104(9):1353-6.
86. Clarke S, Green-Johnson J, Brooks S, Ramdath D, Bercik P, Avila C, Inglis G, Green J, Yanke L, Selinger L, et al. beta-2-1 Fructan supplementation alters host immune responses in a manner consistent with increased exposure to microbial components:

- Results from a double-blinded, randomised, cross-over study in healthy adults. *British journal of nutrition* 2016;115(10):1748-59.
87. Cloetens L, Broekaert WF, Delaedt Y, Ollevier F, Courtin CM, Delcour JA, Rutgeerts P, Verbeke K. Tolerance of arabinoxylan-oligosaccharides and their prebiotic activity in healthy subjects: A randomised, placebo-controlled cross-over study. *British Journal of Nutrition* 2010;103(5):703-13.
  88. Costabile A, Fava F, Röytiö H, Forssten SD, Olli K, Klievink J, Rowland IR, Ouwehand AC, Rastall RA, Gibson GR, et al. Impact of polydextrose on the faecal microbiota: A double-blind, crossover, placebo-controlled feeding study in healthy human subjects. *British Journal of Nutrition* 2012;108(3):471-81.
  89. Costabile A, Klinder A, Fava F, Napolitano A, Fogliano V, Leonard C, Gibson GR, Tuohy KM. Whole-grain wheat breakfast cereal has a prebiotic effect on the human gut microbiota: A double-blind, placebo-controlled, crossover study. *British Journal of Nutrition* 2008;99(1):110-20.
  90. Costabile A, Kolida S, Klinder A, Gietl E, Buerlein M, Froberg C, Landschütze V, Gibson GR. A double-blind, placebo-controlled, cross-over study to establish the bifidogenic effect of a very-long-chain inulin extracted from globe artichoke (*Cynara scolymus*) in healthy human subjects. *British Journal of Nutrition* 2010;104(7):1007-17.
  91. Damen B, Cloetens L, Broekaert WF, François I, Lescroart O, Trogh I, Arnaut F, Welling GW, Wijffels J, Delcour JA, et al. Consumption of breads containing in situ-produced arabinoxylan oligosaccharides alters gastrointestinal effects in healthy volunteers. *Journal of Nutrition* 2012;142(3):470-7.
  92. Depeint F, Tzortzis G, Vulevic J, l'Anson K, Gibson GR. Prebiotic evaluation of a novel galactooligosaccharide mixture produced by the enzymatic activity of

- Bifidobacterium bifidum* NCIMB 41171, in healthy humans: A randomized, double-blind, crossover, placebo-controlled intervention study. *American Journal of Clinical Nutrition* 2008;87(3):785-91.
93. Fernando W, Hill J, Zello G, Tyler R, Dahl W, Kessel A. Diets supplemented with chickpea or its main oligosaccharide component raffinose modify faecal microbial composition in healthy adults. *Beneficial microbes* 2010;1(2):197-207.
  94. François IEJA, Lescroart O, Veraverbeke WS, Marzorati M, Possemiers S, Evenepoel P, Hamer H, Houben E, Windey K, Welling GW, et al. Effects of a wheat bran extract containing arabinoxylan oligosaccharides on gastrointestinal health parameters in healthy adult human volunteers: A double-blind, randomised, placebo-controlled, cross-over trial. *British Journal of Nutrition* 2012;108(12):2229-42.
  95. Fuller Z, Louis P, Mihajlovski A, Rungapamestry V, Ratcliffe B, Duncan AJ. Influence of cabbage processing methods and prebiotic manipulation of colonic microflora on glucosinolate breakdown in man. *British Journal of Nutrition* 2007;98(2):364-72.
  96. Gråsten SM, Juntunen KS, Mättö J, Mykkänen OT, El-Nezami H, Adlercreutz H, Poutanen KS, Mykkänen HM. High-fiber rye bread improves bowel function in postmenopausal women but does not cause other putatively positive changes in the metabolic activity of intestinal microbiota. *Nutrition Research* 2007;27(8):454-61.
  97. Holscher HD, Bauer LL, Gourineni V, Pelkman CL, Fahey GC, Swanson KS. Agave inulin supplementation affects the fecal microbiota of healthy adults participating in a randomized, double-blind, placebo-controlled, crossover trial. *Journal of Nutrition* 2015;145(9):2025-32.
  98. Jenkins DJA, Vuksan V, Rao AV, Vidgen E, Kendall CWC, Tariq N, Würsch P, Koellreutter B, Shiwnarain N, Jeffcoat R. Colonic bacterial activity and serum lipid

- risk factors for cardiovascular disease. *Metabolism: Clinical and Experimental* 1999;48(2):264-8.
99. Maki KC, Gibson GR, Dickmann RS, Kendall CWC, Chen CYO, Costabile A, Comelli EM, McKay DL, Almeida NG, Jenkins D, et al. Digestive and physiologic effects of a wheat bran extract, arabino-xylan-oligosaccharide, in breakfast cereal. *Nutrition* 2012;28(11):1115-21.
  100. Maneerat S, Lehtinen MJ, Childs CE, Forssten SD, Alhoniemi E, Tiphaine M, Yaqoob P, Ouwehand AC, Rastall RA. Consumption of *Bifidobacterium lactis* Bi-07 by healthy elderly adults enhances phagocytic activity of monocytes and granulocytes. *Journal of Nutritional Science* 2013;2:e44. doi: 10.1017/jns.2013.31.
  101. Martínez I, Kim J, Duffy PR, Schlegel VL, Walter J. Resistant starches types 2 and 4 have differential effects on the composition of the fecal microbiota in human subjects. *PLoS ONE* 2010;5(11).
  102. Petry N, Egli I, Chassard C, Lacroix C, Hurrell R. Inulin modifies the bifidobacteria population, fecal lactate concentration, and fecal pH but does not influence iron absorption in women with low iron status. *American Journal of Clinical Nutrition* 2012;96(2):325-31.
  103. Ramnani P, Costabile A, Bustillo AGR, Gibson GR. A randomised, double-blind, cross-over study investigating the prebiotic effect of agave fructans in healthy human subjects. *Journal of Nutritional Science* 2015;4.
  104. Ross AB, Bruce SJ, Blondel-Lubrano A, Oguey-Araymon S, Beaumont M, Bourgeois A, Nielsen-Moennoz C, Vigo M, Fay LB, Kochhar S, et al. A whole-grain cereal-rich diet increases plasma betaine, and tends to decrease total and LDL-cholesterol compared with a refined-grain diet in healthy subjects. *British Journal of Nutrition* 2011;105(10):1492-502.

105. Slavin J, Feirtag J. Chicory inulin does not increase stool weight or speed up intestinal transit time in healthy male subjects. *Food & function* 2011;2(1):72-7.
106. Smith SC, Choy R, Johnson SK, Hall RS, Wildeboer-Veloo ACM, Welling GW. Lupin kernel fiber consumption modifies fecal microbiota in healthy men as determined by rRNA gene fluorescent in situ hybridization. *European Journal of Nutrition* 2006;45(6):335-41.
107. Ten Bruggencate SJM, Bovee-Oudenhoven IMJ, Lettink-Wissink MLG, Katan MB, Van Der Meer R. Dietary fructooligosaccharides affect intestinal barrier function in healthy men. *Journal of Nutrition* 2006;136(1):70-4.
108. Tuohy KM, Kolida S, Lustenberger AM, Gibson GR. The prebiotic effects of biscuits containing partially hydrolysed guar gum and fructo-oligosaccharides - A human volunteer study. *British Journal of Nutrition* 2001;86(3):341-8.
109. Vulevic J, Drakoularakou A, Yaqoob P, Tzortzis G, Gibson GR. Modulation of the fecal microflora profile and immune function by a novel trans-galactooligosaccharide mixture (B-GOS) in healthy elderly volunteers. *American Journal of Clinical Nutrition* 2008;88(5):1438-46.
110. Vulevic J, Juric A, Walton GE, Claus SP, Tzortzis G, Toward RE, Gibson GR. Influence of galacto-oligosaccharide mixture (B-GOS) on gut microbiota, immune parameters and metabonomics in elderly persons. *British Journal of Nutrition* 2015;114(4):586-95.
111. Walton G, Heuvel E, Kusters M, Rastall R, Tuohy K, Gibson G. A randomised crossover study investigating the effects of galacto-oligosaccharides on the faecal microbiota in men and women over 50 years of age. *The British journal of nutrition* 2012;107(10):1466-75.

112. Walton GE, Lu C, Trogh I, Arnaut F, Gibson GR. A randomised, double-blind, placebo controlled cross-over study to determine the gastrointestinal effects of consumption of arabinoxylan- oligosaccharides enriched bread in healthy volunteers. *Nutrition journal* 2012;11(1).
113. Walton GE, Rastall RA, Rastall RA, Martini MC, Williams CE, Jeffries RL, Gibson GR. A double-blind, placebo controlled human study investigating the effects of coffee derived manno-oligosaccharides on the faecal microbiota of a healthy adult population. *International Journal of Probiotics and Prebiotics* 2010;5(2):75-83.
114. Zeng Y, Huang S, Mu G, Zeng X, Zhou X. Effects of whole grain-bean mixed staple food on intestinal microecology and metabolic parameters of obese people. *Chinese Journal of Clinical Nutrition* 2015;23(1):27-34.
115. Blædel T, Holm JB, Sundekilde UK, Schmedes MS, Hess AL, Lorenzen JK, Kristiansen K, Dalsgaard TK, Astrup A, Larsen LH. A randomised, controlled, crossover study of the effect of diet on angiopoietin-like protein 4 (ANGPTL4) through modification of the gut microbiome. *Journal of Science* 2016;5.
116. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, Harris HMB, Coakley M, Lakshminarayanan B, O'Sullivan O, et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature* 2012;488(7410):178-84. doi: <http://www.nature.com/nature/journal/v488/n7410/abs/nature11319.html#supplementary-information>.
117. Coyte KZ, Schluter J, Foster KR. The ecology of the microbiome: Networks, competition, and stability. *Science (New York, NY)* 2015;350(6261):663-6. doi: 10.1126/science.aad2602.
118. Johnson KVA, Burnet PWJ. Microbiome: Should we diversify from diversity? *Gut microbes* 2016;7(6):455-8. doi: 10.1080/19490976.2016.1241933.

119. Van der Meulen R, Adriany T, Verbrugghe K, De Vuyst L. Kinetic analysis of bifidobacterial metabolism reveals a minor role for succinic acid in the regeneration of NAD<sup>+</sup> through its growth-associated production. *Applied and environmental microbiology* 2006;72(8):5204-10. doi: 10.1128/aem.00146-06.
120. Ganzle MG, Follador R. Metabolism of oligosaccharides and starch in lactobacilli: a review. *Front Microbiol* 2012;3:340. doi: 10.3389/fmicb.2012.00340.
121. Gueimonde M, Debor L, Tolkkio S, Jokisalo E, Salminen S. Quantitative assessment of faecal bifidobacterial populations by real-time PCR using lanthanide probes. *Journal of applied microbiology* 2007;102(4):1116-22. doi: 10.1111/j.1365-2672.2006.03145.x.
122. Matsuki T, Watanabe K, Fujimoto J, Miyamoto Y, Takada T, Matsumoto K, Oyaizu H, Tanaka R. Development of 16S rRNA-Gene-Targeted Group-Specific Primers for the Detection and Identification of Predominant Bacteria in Human Feces. *Applied and environmental microbiology* 2002;68(11):5445-51. doi: 10.1128/AEM.68.11.5445-5451.2002.
123. Davis LMG, Martínez I, Walter J, Goin C, Hutkins RW. Barcoded Pyrosequencing Reveals That Consumption of Galactooligosaccharides Results in a Highly Specific Bifidogenic Response in Humans. *PLoS ONE* 2011;6(9):e25200. doi: 10.1371/journal.pone.0025200.
124. Whelan K, Judd PA, Preedy VR, Simmering R, Jann A, Taylor MA. Fructooligosaccharides and fiber partially prevent the alterations in fecal microbiota and short-chain fatty acid concentrations caused by standard enteral formula in healthy humans. *The Journal of nutrition* 2005;135(8):1896-902.
125. Moreno-Indias I, Sanchez-Alcoholado L, Perez-Martinez P, Andres-Lacueva C, Cardona F, Tinahones F, Queipo-Ortuno MI. Red wine polyphenols modulate fecal

- microbiota and reduce markers of the metabolic syndrome in obese patients. *Food Funct* 2016;7(4):1775-87. doi: 10.1039/c5fo00886g.
126. Pokusaeva K, Fitzgerald GF, van Sinderen D. Carbohydrate metabolism in *Bifidobacteria*. *Genes & Nutrition* 2011;6(3):285-306. doi: 10.1007/s12263-010-0206-6.
  127. Canani RB, Costanzo MD, Leone L, Pedata M, Meli R, Calignano A. Potential beneficial effects of butyrate in intestinal and extraintestinal diseases. *World journal of gastroenterology* 2011;17(12):1519-28. doi: 10.3748/wjg.v17.i12. 1519.
  128. den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *Journal of lipid research* 2013;54(9):2325-40. doi: 10.1194/jlr.R036012.



**Table 1:** Statistical analysis for the outcomes reported in  $\geq 2$  randomized controlled trials and included in the meta-analysis.

Outcomes	No. of studies in meta-analysis (references)	$n^I$	Results		Heterogeneity		
			Meta-analysis overall estimate (95% CI)	$P$	Chi-square test	$P$	$I^2$ (%)
<b>Shannon Diversity Index</b>	6 (64, 72, 75, 80, 84, 88)	127	MD: -0.06 (95% CI: -0.25; 0.12)	0.48	10.73	0.06	53
<b>Total number of observed OTUs</b>	3 (72, 75, 84)	53	MD: -4.37 (95% CI: -42.92; 34.19)	0.82	0.07	0.97	0
<b><i>Bifidobacterium</i> spp.</b>	51 (52-58, 60, 61, 63-68, 70-76, 82, 84-94, 96-112, 114)	1629	SMD: 0.64 (95% CI: 0.42; 0.86)	<0.00001	327.93	<0.00001	85
<b><i>Lactobacillus</i> spp.<sup>2</sup></b>	23 (52, 55, 56, 60, 63-65, 67, 68, 73, 75, 76, 84, 87, 93, 96, 97, 99, 104, 105, 107, 111, 114)	670	SMD: 0.22 (95% CI: 0.03; 0.41)	0.02	42.8	0.005	49
<b><i>Faecalibacterium prausnitzii</i></b>	13 (53, 61, 67, 68, 74, 84, 88, 94, 99-101, 110, 112)	519	SMD: 0.14 (95% CI: -0.12; 0.39)	0.29	37.53	0.0002	68
<b><i>Roseburia</i> spp.</b>	4 (68, 79, 84, 97)	189	SMD: 0.33 (95% CI: -0.14; 0.80)	0.17	10.16	0.02	70
<b><i>Eubacterium rectale</i></b>	2 (84, 101)	30	SMD: -0.26 (95% CI: -1.20; 0.67)	0.58	3.94	0.05	75
<b><i>Ruminococcus bromii</i></b>	3 (81, 84, 101)	76	SMD: 0.15 (95% CI: -0.15; 0.45)	0.33	1.1	0.58	0
<b>Total SCFA</b>	13 (52, 55, 59, 63, 64, 67, 73, 80, 82, 84, 86, 91, 94)	406	SMD: 0.11 (95% CI: -0.05; 0.27)	0.19	6.46	0.89	0
<b>Acetate</b>	18 (52, 53, 63, 66, 71, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112)	657	SMD: 0.28 (95% CI: -0.08; 0.63)	0.13	119.36	<0.00001	86
<b>Propionate</b>	19 (52, 53, 63, 66, 71,	677	SMD: 0.01 (95% CI: -0.20; 0.22)	0.95	46.23	0.0003	61

Outcomes	No. of studies in meta-analysis (references)	<i>n</i> <sup>1</sup>	Results		Heterogeneity		
			Meta-analysis overall estimate (95% CI)	<i>P</i>	Chi-square test	<i>P</i>	I <sup>2</sup> (%)
Butyrate	74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112, 115) 20 (52, 53, 59, 63, 66, 71, 74, 77, 80, 82, 84, 86, 90, 91, 93, 94, 96, 103, 112, 115)	712	SMD: 0.24 (95% CI: 0.00; 0.47)	0.05	64.21	<0.00001	70

Data was meta-analyzed using a random-effects model and presented as MDs or SMDs as appropriate. Statistical heterogeneity was assessed using the chi-square test and quantified using the I<sup>2</sup> statistic. <sup>1</sup> Number of participants in meta-analysis. <sup>2</sup> Results from outlier study excluded from this meta-analysis. Abbreviations: MD, Mean difference; OTU, Operational taxonomic unit; SCFA, Short chain fatty acid; SMD, Standardized mean difference.

**Table 2:** Characteristics of randomized controlled trials of fiber supplementation comparing dietary fiber with placebo or low fiber comparators in healthy adults

Study	Participants		Interventions			RCT Design				
	n; age <sup>1</sup> ; % F	Fiber, daily dose	Prebiotic	Comparator; daily dose	Compliance <sup>2</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Abell 2008 (81)</b>	46; 25-66; 65%	RS, 22 g	C	RS, 1 g	Y	Cross-over	28	Y	Y	qPCR
<b>Alfa 2017 (77)</b>	84; 32-96; 42%	RS2, 21 g	C	Corn starch, 21 g	Y	Parallel	72	Y	N	Illumina
<b>Alles 1999 (52)</b>	27.4; 40.4; 45%	TOS, 15 g	A	Glucose & lactose mix, 15 g	Y	Parallel	21	Y	N	Culture
<b>Baer 2014 (83)</b>	14; 47; 9%	Resistant maltodextrin, 50 g	C	Maltodextrin, 50 g	Y	Cross-over	21	N	Y	454 Pyrosequencing; DGGE; FISH; qPCR
<b>Beards 2010 (53)</b>	30; 33 <sup>3</sup> ; 66% <sup>3</sup>	PDX; RS, 45.6 g	C	Maltitol, 45.6 g	N	Parallel	44	N	N	FISH
<b>Blaedel 2016 (115)</b>	21; 23-45; 100%	Inulin, 15 g	A	Placebo	Y	Cross-over	21	N	Y	Illumina
<b>Boler 2011 (84); Hooda 2012 (49)<sup>4</sup></b>	21; 21-28; 0%	<b>PDX<sup>5</sup></b> ; Soluble maize fiber, 21 g	C	Placebo	N	Cross-over	21	N	N	qPCR; Pyrosequencing <sup>4</sup>
<b>Bouhnik 1996 (54)</b>	10; 22-39; 50%	SC-FOS, 12.5 g	A	Saccharose, 10 g	N	Parallel	12	Y	Y	Culture
<b>Bouhnik 1999 (58)</b>	8; 29.6; 55%	SC-FOS, 20 g	A	Saccharose, 20 g	N	Parallel	7	N	N	Culture

Study	Participants		Interventions			RCT Design				
	n; age <sup>1</sup> ; % F	Fiber, daily dose	Prebiotic	Comparator; daily dose	Compliance <sup>2</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Bouhnik 2004 (57)</b>	64; 30 <sup>3</sup> ; 55% <sup>3</sup>	SC-FOS <sup>5</sup> ; GOS <sup>5</sup> ; Isomalto-OS; <b>Inulin</b> <sup>5</sup> ; RS; Soybean-OS, 10 g	A	Sucrose & maltodextrin mix, 10 g	N	Parallel	7	Y	N	Culture
<b>Bouhnik 2006 (56)</b>	40; 29; 55%	SC-FOS (Actilight), 10 g	A	Sucrose & maltodextrin mix, 10 g	N	Parallel	7	Y	N	Culture
<b>Bouhnik 2007 (55)</b>	39; 33.9; NR	Inulin, 5 g	A	Sucrose & maltodextrin mix, 5 g	N	Parallel	28	Y	Y	Culture
<b>Brahe 2015 (59)</b>	35; 59.6 <sup>3</sup> ; 100%	Flaxseed mucilage, 10 g	G	Placebo	Y	Parallel	42	N	N	Quantitative metagenomics qPCR
<b>Calame 2008 (60)</b>	16; 30.9; NR	Arabic gum, 40 g	G	Placebo	Y	Parallel	28	N	N	
<b>Clarke 2016 (86)</b>	30; 27; 57%	Beta 2-1 fructan, 15 g	A	Maltodextrin, 15 g	Y	Cross-over	28	N	Y	qPCR
<b>Cloetens 2010 (87)</b>	20; 24; 70%	AXOS, 10 g	C	Maltodextrin, 20 g	N	Cross-over	21	N	Y	qPCR
<b>Costabile 2010 (90)</b>	31; 25; 56%	Very long chain inulin, 10 g	A	Maltodextrin, 10 g	N	Cross-over	21	N	Y	FISH
<b>Costabile 2012 (88)</b>	31; 33; 52%	PDX, 8 g	C	Maltodextrin, 8 g	N	Cross-over	21	N	Y	DGGE; FISH
<b>Damen 2012 (91)</b>	27; 25; 63%	AXOS, 2.14 g	C	Placebo	Y	Cross-over	21	Y	Y	FISH
<b>Depeint 2008 (92)</b>	30; 36.3; 60%	Beta-GOS, 7 g	A	Sucrose, 7 g	N	Cross-over	7	Y	Y	FISH

Study	Participants		Interventions			RCT Design				
	n; age <sup>1</sup> ; % F	Fiber, daily dose	Prebiotic	Comparator; daily dose	Compliance <sup>2</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Dewulf 2013 (61)</b>	30; 47.5; 100%	Inulin-type fructan (Synergy 1), 16 g	A	Maltodextrin, 16 g	N	Parallel	Reported as 3 months	N	N	qPCR; Phylogenetic microarray
<b>Elison 2016 (62)</b>	40; 22-57; 52%	<b>HMO<sup>6</sup></b> : 2'-O-fucosyllactose (2'FL); lacto-N-neotetraose (LNnT); Mixture (2:1 mixture of 2'FL + LNnT), 20 g	A	Glucose, 2 g	Y	Parallel	14	Y	N	Illumina
<b>Fastinger 2008 (63)</b>	25; 26.7; 50%	Resistant maltodextrin, 15 g	C	Maltodextrin, 15 g	N	Parallel	21	Y	Y	qPCR
<b>Fernando 2010 (93)</b>	12; 25.6; 42%	Raffinose, 5 g	G	Placebo	N	Cross-over	21	N	N	qPCR; T-RLFP
<b>Finegold 2014 (64)</b>	16; 21-49 <sup>3</sup> ; 66% <sup>3</sup>	XOS, 2.8 g	C	Maltodextrin, 2.8 g	N	Parallel	56	Y	Y	Pyrosequencing
<b>Francois 2012 (94)</b>	52; 42; 48%	Wheat bran extract, 10 g	G	Placebo	N	Cross-over	21	Y	Y	FISH
<b>Fuller 2007 (95); Ramirez-Farias 2009 (50)<sup>4</sup></b>	12; 38.1; 75%	Inulin, 10 g	A	Nil	Y	Cross-over	16	N	N	qPCR
<b>Gopal 2003 (65)</b>	19; 20-60 <sup>3</sup> ; 44% <sup>3</sup>	GOS, 2.4 g	A	Placebo	Y	Parallel	28	Y	Y	Culture
<b>Holscher 2015 (97)</b>	29; 27; 52%	Agave inulin, 7.5 g	A	Placebo	N	Cross-over	21	Y	Y	Illumina

Study	Participants		Interventions			RCT Design				
	n; age <sup>1</sup> ; % F	Fiber, daily dose	Prebiotic	Comparator; daily dose	Compliance <sup>2</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Jie 2000 (66)</b>	30; 29.9; 45%	PDX, 12 g	C	Nil	N	Parallel	28	Y	N	Culture
<b>Kleesen 2007 (67)</b>	45; 23.5; 55%	<b>Inulin</b> <sup>6</sup> : Chicory inulin; Jerusalem artichoke inulin, 15.4 g	A	Placebo	N	Parallel	21	Y	N	Culture; FISH
<b>Lecerf 2012 (68)</b>	59; 20.1; 57%	<b>XOS</b> <sup>5</sup> ; Inulin-XOS mix, 6.64 g	C	Wheat dextrin, 6.64 g	N	Parallel	28	N	N	qPCR
<b>Lin 2016 (69)</b>	20; 24.2; 80%	XOS, 1.2 g	C	Placebo	N	Parallel	42	Y	Y	Culture
<b>Lomax 2012 (70)</b>	43; 55; 74%	Beta 2-1 fructan, 8 g	A	Maltodextrin, 8 g	Y	Parallel	28	Y	N	FISH
<b>Maki 2012 (99)</b>	55; 35.1 <sup>3</sup> ; 54% <sup>3</sup>	AXOS, 2.4 g	C	Placebo	N	Cross-over	21	N	Y	FISH
<b>Maneerat 2013 (100)</b>	35; 67.4 <sup>3</sup> ; 53% <sup>3</sup>	GOS, 8 g	A	Maltodextrin, 8 g	N	Cross-over	21	N	Y	FISH
<b>Martinez 2010 (101)</b>	10; 23-38; 50%	<b>RS</b> <sup>6</sup> : RS2; RS4, 33.2 g	C	Native wheat starch, 33.2 g	N	Cross-over	21	Y	Y	Pyrosequencing
<b>Pallav 2014 (72)</b>	14; 31.4 <sup>3</sup> ; 65%	Polysaccharidepeptide (I'm-Yunity), 3.6 g	G	Nil	N	Parallel	14	N	N	Pyrosequencing
<b>Pasman 2006 (73)</b>	29; 34.1; 0%	Nutriose FB (dextrin), 45 g	A	Maltodextrin, 22.5 g	Y	Parallel	35	Y	N	Culture
<b>Petry 2012 (102)</b>	32; 18-40; 100%	Inulin, 20 g	A	Maltodextrin, 20 g	N	Cross-over	28	N	Y	qPCR
<b>Ramnani 2010 (74)</b>	66; 32.9; 50%	Inulin, 5 g	A	Placebo	Y	Parallel	21	Y	Y	FISH

Study	Participants		Interventions			RCT Design				
	n; age <sup>1</sup> ; % F	Fiber, daily dose	Prebiotic	Comparator; daily dose	Compliance <sup>2</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Ramnani 2015 (103)</b>	38; 35.1 <sup>3</sup> ; 50%	Agave inulin, 5 g	A	Maltodextrin, 5 g	Y	Cross-over	21	Y	Y	FISH
<b>Salden 2017 (80)</b>	27; 48; 48%	Arabinoxylans, 15 g	G	Maltodextrin, 15 g	Y	Parallel	42	N	N	Illumina
<b>Slavin 2011 (105)</b>	10; 27-49 <sup>3</sup> ; 0%	Chicory inulin, 20 g	A	Placebo	Y	Cross-over	21	N	N	Culture
<b>Ten Bruggenca 2006 (107)</b>	29; 22.7; 0%	FOS, 20 g	A	Sucrose, 6 g	Y	Cross-over	14	N	Y	qPCR
<b>Tuohy 2011 (108)</b>	NR; NR; 55%	Mix:(FOS & PHGG), 10 g	Mix	Placebo	Y	Cross-over	21	N	N	FISH
<b>Vulevic 2008 (109)</b>	41; 69.3 <sup>3</sup> ; 64% <sup>3</sup>	GOS (Bimuno), 5.5 g	A	Maltodextrin, 5.5 g	Y	Cross-over	70	N	Y	FISH
<b>Vulevic 2015 (110)</b>	40; 70.4; 62%	GOS (Bimuno), 5.5 g	A	Maltodextrin, 5.5 g	Y	Cross-over	70	N	Y	FISH
<b>Walton 2010 (113)</b>	31; 21; 58%	MOS, 5 g	C	Placebo	Y	Cross-over	21	N	Y	FISH
<b>Walton 2012 (111)</b>	37; 58.9 <sup>3</sup> ; 57% <sup>3</sup>	GOS, 8 g	A	Placebo	N	Cross-over	21	Y	Y	qPCR
<b>Walton 2012 (112)</b>	40; 31.4 <sup>3</sup> ; 60% <sup>3</sup>	AXOS, 2.2 g	C	Placebo	Y	Cross-over	21	Y	Y	FISH
<b>Wu 2011 (76)</b>	15; 40.6; 93%	Konjac glucomannan, 4.5 g	G	Nil	N	Parallel	28	N	N	FISH

<sup>1</sup> Age expressed as mean years; age range provided where means were not obtainable. <sup>2</sup> Compliance to intervention; assessed by primary study. <sup>3</sup> Refers to randomized population rather than actual population. Compliance to intervention; assessed by primary study. <sup>4</sup> Secondary publication reporting additional outcomes from the primary study. <sup>5</sup> Refers to analyzed intervention arm with the highest prebiotic classification (accepted prebiotic fiber > candidate prebiotic fiber > general fiber) selected for fiber type subgroup analysis. <sup>6</sup> Refers to intervention fibers that have been pooled together for meta-analyses. Abbreviations: A; Accepted prebiotic fiber; AXOS; Arabinoxylan-oligosaccharide; C; Candidate prebiotic fiber; DGGE; Denaturing gradient gel electrophoresis; FISH; Fluorescent *in situ* hybridization; G; General fiber; GOS; Galacto-oligosaccharide; HMO; Human milk oligosaccharide; MOS; Manno-oligosaccharide; NR; Not reported by study; OS; Oligosaccharide; PDX; Polydextrose; PHGG; Partially hydrolyzed guar gum; qPCR; Quantitative polymerase chain reaction; RS; Resistant starch; RS2; Resistant starch 2; RS4; Resistant starch 4; SC-FOS; Short chain fructo-oligosaccharide; TOS; Trans-galacto-oligosaccharide; XOS; Xylo-oligosaccharide.



**Table 3:** Characteristics of randomized controlled trials of food interventions comparing dietary fiber with low fiber comparators in healthy adults

Study	Participants	Interventions			RCT Design						
	n; age <sup>1</sup> ; % F	Intervention	Comparator	Daily fiber difference	Study diet <sup>2</sup>	Compliance <sup>3</sup>	Design	Duration (days)	Run in	Wash out	Analysis
<b>Ampatzoglou 2008 (82)</b>	33; 48.8; 64%	WG diet	RG diet	10 g	N	Y	Cross-over	14	Y	Y	FISH
<b>Carvalho-Wells 2010 (85)</b>	32; 31.6; 66%	WG cereal	Non-WG cereal	6.5 g	N	N	Cross-over	21	Y	Y	FISH
<b>Cooper 2017 (78)</b>	46; 25.8; 46%	WG market basket	RG market basket	5 g	N	Y	Parallel	42	N	N	Illumina
<b>Costabile 2008 (89)</b>	31; 25; 52%	WG cereal	Wheat bran cereal	7.4 g	N	N	Cross-over	21	Y	Y	FISH
<b>Grasten 2007 (96)</b>	14; 59.7 <sup>4</sup> ; 100%	Rye bread	White wheat bread	19 g	N	Y	Cross-over	56	Y	Y	Culture
<b>Jenkins 1999 (98)</b>	24; 33; 50%	Wheat bran	Wheat flour	19 g	N	Y	Cross-over	14	N	Y	Culture
<b>Karl 2017 (79); Vanegas 2017 (51)<sup>5</sup></b>	81; 40-65 <sup>4</sup> ; 60%	WG diet	RG diet	8 g	Y	Y	Parallel	42	Y	N	Illumina
<b>Nemoto 2011 (71)</b>	36; 22-67; 63%	Fermented brown rice	"Non-functional food"	4.62 g	N	Y	Parallel	14	N	N	Culture
<b>Ross 2011</b>	17; 35; 65%	WG diet	RG diet	13 g	Y	Y	Cross-	14	Y	Y	qPCR

Study	Participants	Interventions			RCT Design						
	n; age <sup>1</sup> ; % F	Intervention	Comparator	Daily fiber difference	Study diet <sup>2</sup>	Compliance <sup>3</sup>	Design	Duration (days)	Run in	Wash out	Analysis
(104)							over				
Smith 2006 (106)	18; 42.8; 0%	Lupin kernal fiber diet	Control diet	22 g	Y	N	Cross-over	28	N	Y	FISH
Tap 2015 (75)	19; 19-25; 53%	High fiber diet	Low fiber diet	30 g	Y	Y	Cross-over	5	N	Y	454 Pyrosequencing
Zeng 2015 (114)	77; 63.4; 70%	Whole cereal legume diet	Control diet	14.5 g	Y	Y	Parallel	90	N	N	Culture

<sup>1</sup> Age expressed as mean years; age range provided where means were not obtainable. <sup>2</sup> Whether the participant's entire diet was provided by the study. <sup>3</sup> Compliance to intervention; assessed by primary study. <sup>4</sup> Refers to randomized population rather than actual population. <sup>5</sup> Secondary publication reporting additional outcomes from the primary study. Abbreviations: FISH; Fluorescent *in situ* hybridization; qPCR; Quantitative polymerase chain reaction; RG; Refined grain; WG; Whole grain.

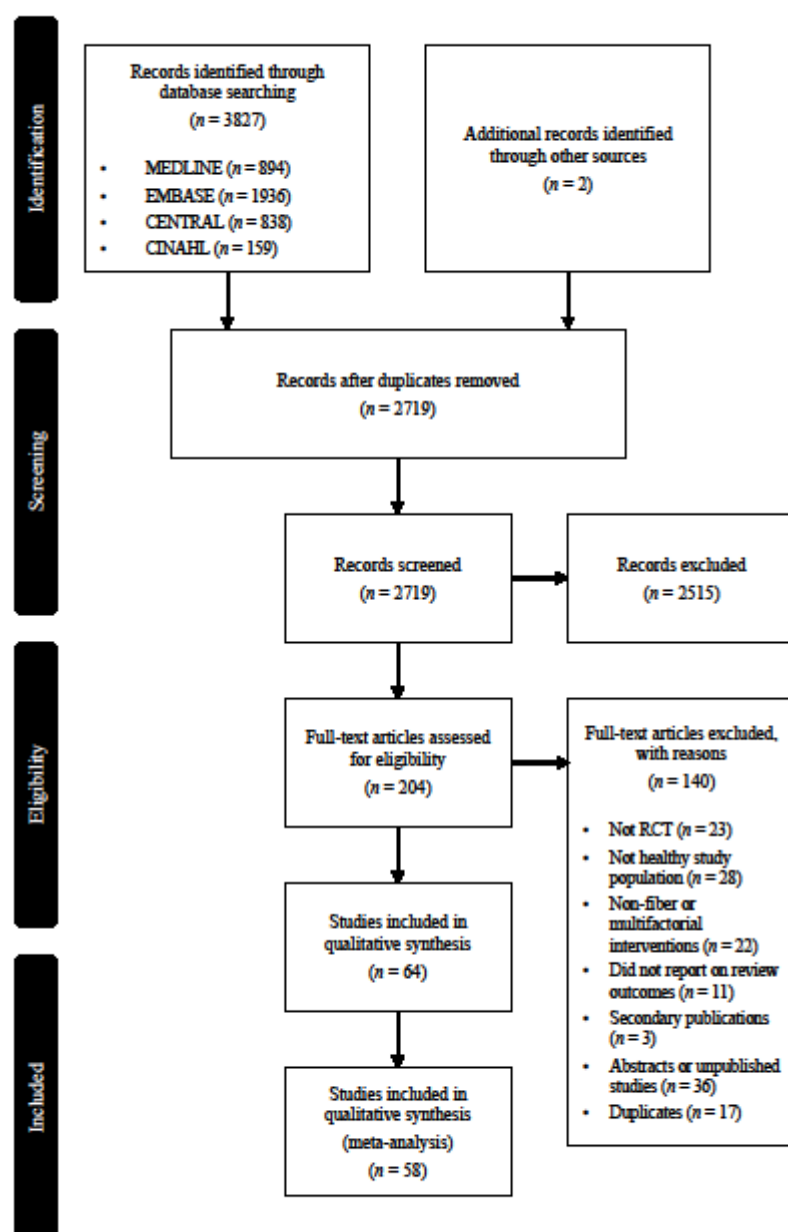


Figure 1: Flow diagram of studies evaluated in the systematic review.

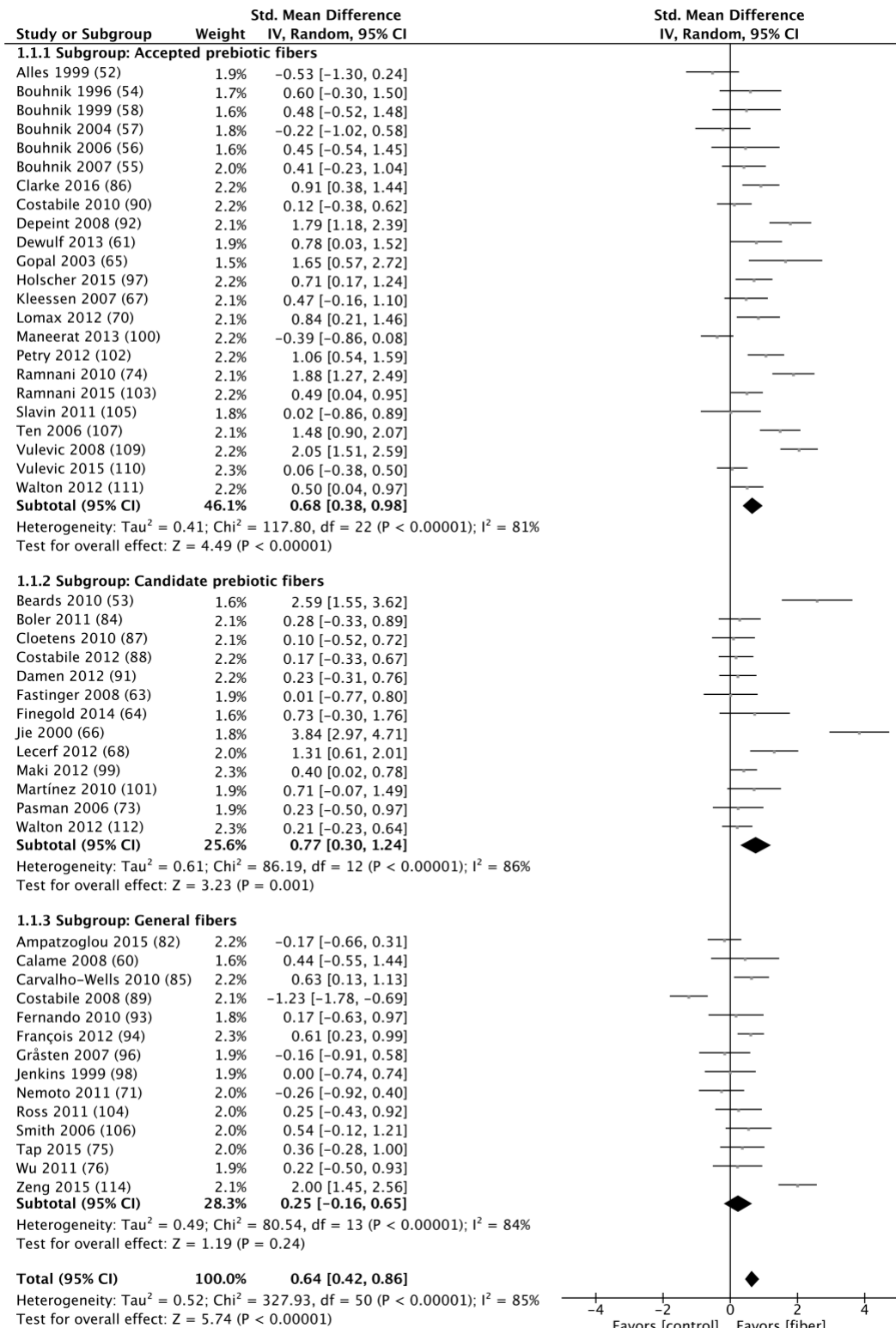


Figure 2: Forest plot of randomized controlled trials in healthy adults comparing dietary fiber with placebo/low fiber comparators. Studies are sub-grouped by fiber type, with the overall effect included at the bottom. Data are presented as means and SDs of *Bifidobacterium* spp. abundance at end of intervention. Effects of trials are presented as weights (percentages) and SMD (95% CI). CI, confidence interval; IV, inverse variance; SD, standard deviation; SMD, standardized mean difference.

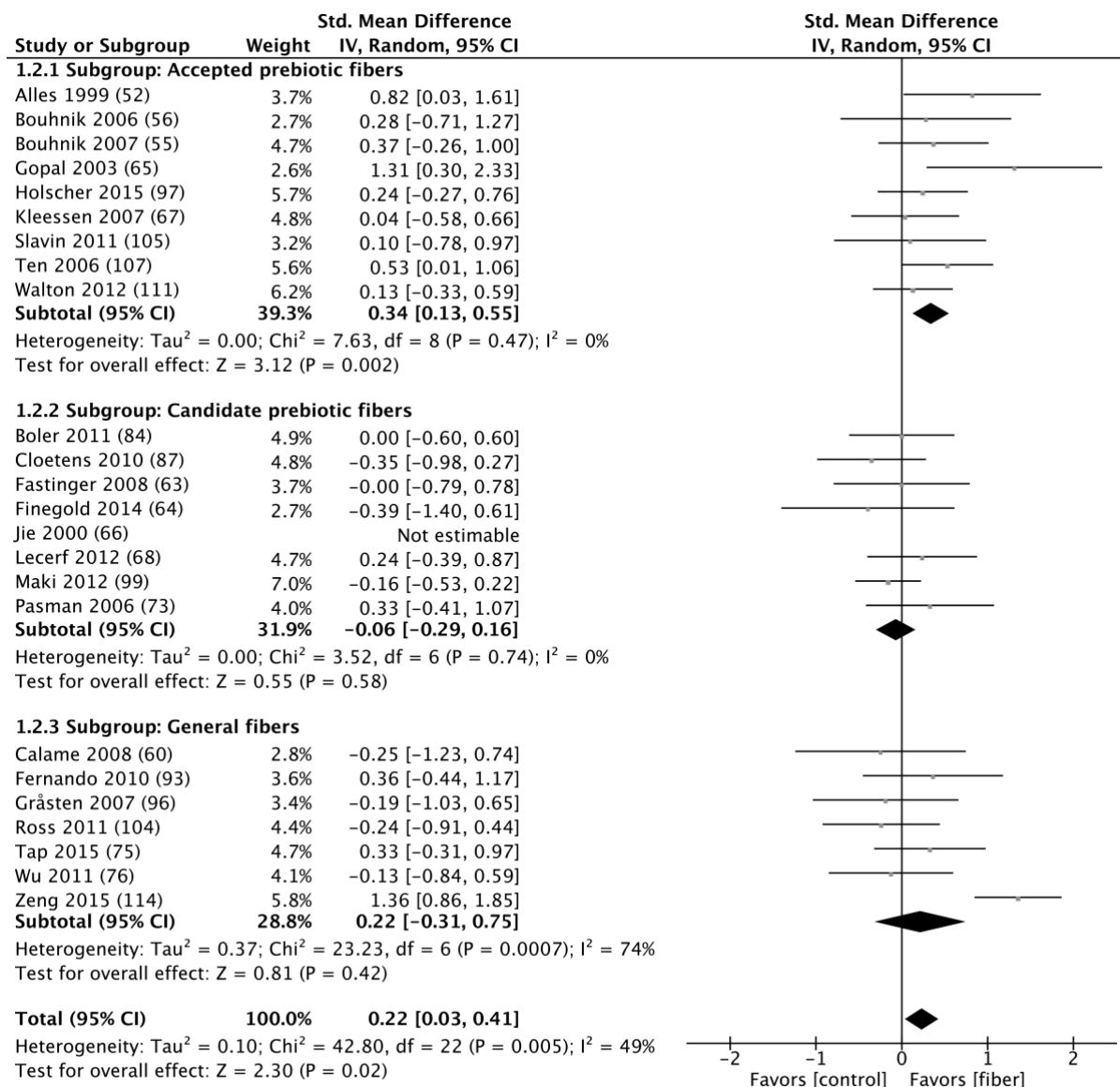


Figure 3: Forest plot of randomized controlled trials in healthy adults comparing dietary fiber with placebo/low fiber comparators. Studies are sub-grouped by fiber type, with the overall effect included at the bottom. Data are presented as means and SDs of *Lactobacillus* spp. abundance at end of intervention are reported for trials. Effects of trials are presented as weights (percentages) and SMD (95% CI). CI, confidence interval; IV, inverse variance; SD, standard deviation; SMD, standardized mean difference.

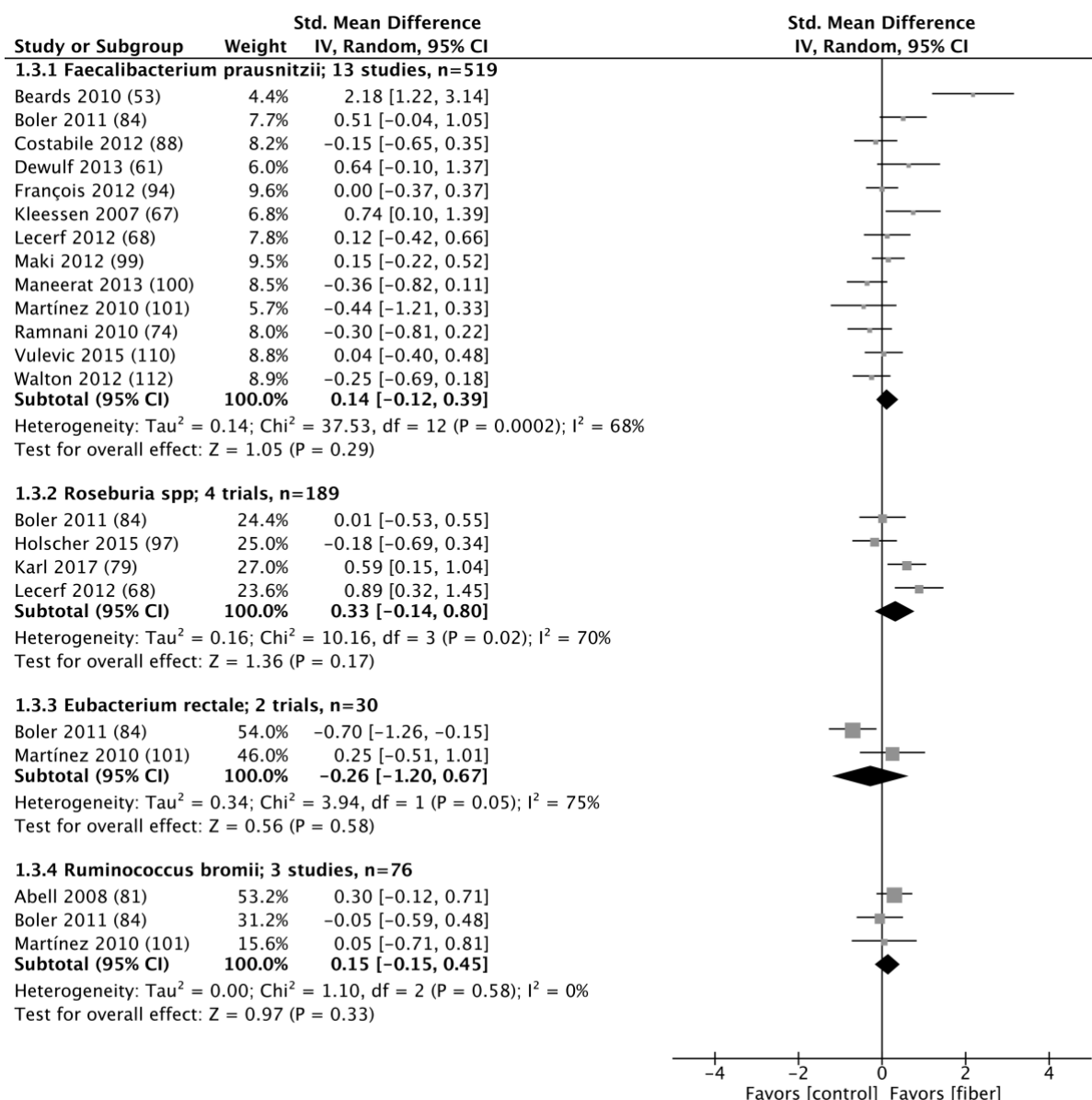


Figure 4: Forest plot of randomized controlled trials in healthy adults comparing dietary fiber with placebo/low fiber comparators. The means and SDs of *Faecalibacterium prausnitzii*, *Roseburia* spp., *Eubacterium rectale* and *Ruminococcus bromii* abundance at end of intervention are reported for trials. Effects of trials are presented as weights (percentages) and SMD (95% CI). CI, confidence interval; IV, inverse variance; SD, standard deviation; SMD, standardized mean difference.

Supplemental Table 1: Search algorithm: MEDLINE via OVID

Supplemental Table 2: Search algorithm: EMBASE

Supplemental Table 3: Search algorithm: CENTRAL

Supplemental Table 4: Search algorithm: CINAHL

Supplemental Table 5: Reasons for excluding studies from full text analysis

Supplemental Table 6: Outcomes of pre-defined subgroup analyses undertaken

Supplemental Table 7: Outcomes of post hoc subgroup analyses undertaken

Supplemental Figure 1: Risk of bias across the included studies showing the summary percentage in each domain

Supplemental Figure 2: Funnel plot for the effect of dietary fiber on *Bifidobacterium* spp. abundance

Supplemental Figure 3: Funnel plot for the effect of dietary fiber on *Lactobacillus* spp. abundance

Supplemental Figure 4: Funnel plot for the effect of dietary fiber on total fecal SCFA

Supplemental Figure 5: Funnel plot for the effect of dietary fiber on fecal acetate

Supplemental Figure 6: Funnel plot for the effect of dietary fiber on fecal propionate

Supplemental Figure 7: Funnel plot for the effect of dietary fiber on fecal butyrate

**Supplemental Table 1:** Search algorithm: MEDLINE via OVID

1. exp Dietary Fiber/	46. exp Inulin/
2. roughage*.tw.	47. Inulin*.tw.
3. exp Prebiotics/	48. (gentiooligosaccharide* or gentio-
4. prebiotic*.tw.	oligosaccharide*).tw.
5. (carbohydrate adj2 polymer*).tw.	49. (isomalto oligosaccharide* or isomalto-
6. ((non-starch or nonstarch) adj (poly-saccharide* or	oligosaccharide* or imo).tw.
polysaccharide*)).tw.	50. (mannanooligosaccharide* or mannano-
7. 1 or 2 or 3 or 4 or 5 or 6	oligosaccharide*).tw.
8. Diet/	51. (N-acetylchitooligosaccharide* or N-acetylchito-
9. diet*.tw.	oligosaccharide*).tw.
10. consum*.tw.	52. (pectic oligosaccharide* or pectic-
11. eat*.tw.	oligosaccharide*).tw.
12. food*.tw.	53. (resistant starch* or resistant-starch*).tw.
13. nutri*.tw.	
14. 8 or 9 or 10 or 11 or 12 or 13	54. (soybean oligosaccharide* or soybean-
15. Agar/	oligosaccharide*).tw.
16. agar*.tw.	55. (xylooligosaccharide* or xylo-
17. Alginates/	oligosaccharide*).tw.
18. alginate*.tw.	56. exp Oligosaccharides/
19. (alginic adj2 acid*).tw.	57. Oligosaccharide*.tw.
20. Carrageenan/	58. (fiber* or fiber* or high-fiber* or high-fiber*).tw.
21. carrageen*.tw.	59. 15 or 16 or 17 or 18 or 19 or 20 or 21 or 22 or 23
22. exp Cellulose/	or 24 or 25 or 26 or 27 or 28 or 29 or 30 or 31 or 32
23. cellulose*.tw.	or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41
24. exp Chitin/	or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50
25. chitin*.tw.	or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58
26. hemicellulose*.tw.	
27. hexosan*.tw.	60. 14 and 59
28. Lignin/	61. 7 or 60
29. lignin*.tw.	62. exp Gastrointestinal Microbiome/
30. Pectins/	63. (microbiota or microbiome).tw.
31. pectin*.tw.	64. bifido*.tw.
32. pentosan*.tw.	65. lactobacill*.tw.
33. polydextrose*.tw.	66. 62 or 63 or 64 or 65
34. polyuronide*.tw.	67. (faecal or fecal).tw.
35. Raffinose/	68. (bacteri* or flora).tw.
36. raffinose*.tw.	69. 67 and 68
37. xanthan*.tw.	70. exp Dysbiosis/
38. Xylose/	71. 66 or 69 or 70
39. xylose*.tw.	72. 61 and 71
40. exp Galactans/	73. ((randomized controlled trial or controlled
41. galactan*.tw.	clinical trial).pt. or randomized.ab. or randomised.ab.
42. (galactooligosaccharide* or galacto-	or placebo.ab. or drug therapy.fs. or randomly.ab. or
oligosaccharide* or gos or tos).tw.	trial.ab. or groups.ab.) not (exp animals/ not
43. exp Fructans/	humans.sh.)
44. fructan*.tw.	74. 72 and 73
45. (fructooligosaccharide* or fructo-	
oligosaccharide* or fos or oligofructose or oligo-	
fructose).tw.	



**Supplemental Table 2:** Search algorithm: EMBASE

1. exp Dietary Fiber/	46. exp Inulin/
2. roughage*.tw.	47. Inulin*.tw.
3. exp Prebiotics/	48. (gentiooligosaccharide* or gentio-
4. prebiotic*.tw.	oligosaccharide*).tw.
5. (carbohydrate adj2 polymer*).tw.	49. (isomalto oligosaccharide* or isomalto-
6. ((non-starch or nonstarch) adj (poly-saccharide* or	oligosaccharide* or imo).tw.
polysaccharide*)).tw.	50. (mannanoligosaccharide* or mannano-
7. 1 or 2 or 3 or 4 or 5 or 6	oligosaccharide*).tw.
8. Diet/	51. (N-acetylchitooligosaccharide* or N-acetylchito-
9. diet*.tw.	oligosaccharide*).tw.
10. consum*.tw.	52. (pectic oligosaccharide* or pectic-
11. eat*.tw.	oligosaccharide*).tw.
12. food*.tw.	53. (resistant starch* or resistant-starch*).tw.
13. nutri*.tw.	54. (soybean oligosaccharide* or soybean-
14. 8 or 9 or 10 or 11 or 12 or 13	oligosaccharide*).tw.
15. Agar/	55. (xylooligosaccharide* or xylo-
16. agar*.tw.	oligosaccharide*).tw.
17. Alginates/	56. exp Oligosaccharides/
18. alginate*.tw.	57. Oligosaccharide*.tw.
19. (alginic adj2 acid*).tw.	58. (fiber* or fiber* or high-fiber* or high-fiber*).tw.
20. Carrageenan/	59. 15 or 16 or 17 or 18 or 19 or 20 or 21 or 22 or 23
21. carrageen*.tw.	or 24 or 25 or 26 or 27 or 28 or 29 or 30 or 31 or 32
22. exp Cellulose/	or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41
23. cellulose*.tw.	or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50
24. exp Chitin/	or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58
25. chitin*.tw.	
26. hemicellulose*.tw.	60. 14 and 59
27. hexosan*.tw.	61. 7 or 60
28. Lignin/	62. exp Gastrointestinal Microbiome/
29. lignin*.tw.	63. (microbiota or microbiome).tw.
30. Pectins/	64. bifido*.tw.
31. pectin*.tw.	65. lactobacill*.tw.
32. pentosan*.tw.	66. 62 or 63 or 64 or 65
33. polydextrose*.tw.	67. (faecal or fecal).tw.
34. polyuronide*.tw.	68. (bacteri* or flora).tw.
35. Raffinose/	69. 67 and 68
36. raffinose*.tw.	70. exp Dysbiosis/
37. xanthan*.tw.	71. 66 or 69 or 70
38. Xylose/	72. 61 and 71
39. xylose*.tw.	73. ((randomized controlled trial or controlled
40. exp Galactans/	clinical trial).pt. or randomized.ab. or randomised.ab.
41. galactan*.tw.	or placebo.ab. or drug therapy.fs. or randomly.ab. or
42. (galactooligosaccharide* or galacto-	trial.ab. or groups.ab.) not (exp animals/ not
oligosaccharide* or gos or tos).tw.	humans.sh.)
43. exp Fructans/	74. 72 and 73
44. fructan*.tw.	
45. (fructooligosaccharide* or fructo-	
oligosaccharide* or fos or oligofructose or oligo-	
fructose).tw.	

**Supplemental Table 3:** Search algorithm: CENTRAL

#1	MeSH descriptor: [Dietary Fiber]	#40	MeSH descriptor: [Galactans] explode all trees
explode all trees		#41	galactan*
#2	roughage*	#42	(galactooligosaccharide* or galacto-
#3	MeSH descriptor: [Prebiotics] explode	oligosaccharide* or gos or tos)	
all trees		#43	MeSH descriptor: [Fructans] explode all trees
#4	prebiotic*	#44	fructan*
#5	carbohydrate near/2 polymer*	#45	(fructooligosaccharide* or fructo-
#6	((non-starch or nonstarch) near (poly-	oligosaccharide* or fos or oligofructose* or oligo-	
saccharide* or polysaccharide*))		fructose*)	
#7	#1 or #2 or #3 or #4 or #5 or #6	#46	MeSH descriptor: [Inulin] explode all trees
#8	MeSH descriptor: [Diet] this term only	#47	inulin*
#9	diet*	#48	(gentiooligosaccharide* or gentio-
#10	consum*	oligosaccharide*)	
#11	eat*	#49	(isomalto oligosaccharide* or isomalto-
#12	food*	oligosaccharide* or imo)	
#13	nutri*	#50	(mannanooligosaccharide* or mannano-
#14	#8 or #9 or #10 or #11 or #12 or #13	oligosaccharide*)	
#15	MeSH descriptor: [Agar] this term	#51	(N-acetylchitooligosaccharide* or N-acetylchito-
only		oligosaccharide*)	
#16	agar*	#52	(pectic oligosaccharide* or pectic-
#17	MeSH descriptor: [Alginates] this term	oligosaccharide*)	
only		#53	(resistant starch* or resistant-starch*)
#18	alginate*	#54	(soybean oligosaccharide* or soybean-
#19	alginic near/2 acid	oligosaccharide*)	
#20	MeSH descriptor: [Carrageenan] this	#55	(xylooligosaccharide* or xylo-oligosaccharide*)
term only		#56	MeSH descriptor: [Oligosaccharides] explode all
#21	carrageen*	trees	
#22	MeSH descriptor: [Cellulose] explode	#57	oligosaccharide*
all trees		#58	fiber* or fiber* or high-fiber* or high-fiber*
#23	cellulose*	#59	#15 or #16 or #17 or #18 or #19 or #20 or #21 or
#24	MeSH descriptor: [Chitin] explode all	#22 or #23 or #24 or #25 or #26 or #27 or #28 or #29 or	
trees		#30 or #31 or #32 or #33 or #34 or #35 or #36 or #37 or	
#25	chitin*	#38 or #39 or #40 or #41 or #42 or #43 or #44 or #45 or	
#26	hemicellulose*	#46 or #47 or #48 or #49 or #50 or #51 or #51 or #52 or	
#27	hexosan*	#53 or #54 or #55 or #56 or #56 or #57 or #58	
#28	MeSH descriptor: [Lignin] this term	#60	#14 and #59
only		#61	#7 or #60
#29	lignin*	#62	MeSH descriptor: [Gastrointestinal Microbiome]
#30	MeSH descriptor: [Pectins] this term	explode all trees	
only		#63	(microbiota or microbiome)
#31	pectin*	#64	bifido*
#32	pentosan*	#65	lactobacill*
#33	polydextrose*	#66	#62 or #63 or #64 or #65
#34	polyuronide*	#67	(faecal or fecal)
#35	MeSH descriptor: [Raffinose] this term	#68	(bacteri* or flora)
only		#69	#67 and #68
#36	raffinose*	#70	MeSH descriptor: [Dysbiosis] explode all trees
#37	xanthan*	#71	#66 or #69 or #70
#38	MeSH descriptor: [Xylose] this term	#72	#61 and #71
only			
#39	xylose*		

**Supplemental Table 4:** Search algorithm: CINAHL

1. ((dietary fib\* OR roughage\* OR prebiotic\*) OR (diet\* OR consum\* OR eat\* OR food\* OR nutri\*) AND (agar\* OR alginate\* OR carrageen\* OR cellulose\* OR chitin\* OR hemicellulose\* OR hexosan\* OR lignin\* OR pectin\* OR pentosan\* OR polydextrose\* OR polyuronide\* OR raffinose\* OR xanthan\* OR xylose\* OR galactan\* OR galactooligosaccharde\* OR galacto-oligosaccharide\* OR gos OR tos OR fructan\* OR fructooligosaccharide\* OR fructo-oligosaccharide\* OR fos OR oligofructose\* OR oligo-fructose\* OR inulin\* OR gentiooligosaccharide\* OR gentio-oligosaccharide\* OR isomalto oligosaccharide\* OR isomalto-oligosaccharide\* OR imo OR mannanooligosaccharide\* OR mannano-oligosaccharide\* OR N-acetylchitooligosaccharide\* OR N-acetylchito-oligosaccharide\* OR pectic oligosaccharide\* OR pectic-oigosaccharide\* OR resistant starch\* OR resistant-starch\* OR soybean oligosaccharide\* OR soybean-oligosaccharide\* OR oligosaccharide\* OR high-fib\*))
2. ((MH "Microbiota") OR microbiota OR microbiome OR bifido\* OR lactobacill\*) OR ((faecal OR fecal) AND (bacteri\* OR flora)) OR (dysbio\*)
3. (MH "Clinical Trials+") OR (MH "Quantitative Studies") OR TI placebo\* OR AB placebo\* OR (MH "Placebos") OR (MH "Random Assignment") OR TI random\* OR AB random\* OR TI ((singl\* or doubl\* or tripl\* or trebl\*) W1 (blind\* or mask\*)) OR AB ((singl\* or doubl\* or tripl\* or trebl\*) W1 (blind\* or mask\*)) OR TI clinic\* trial\* OR AB clinic\* trial\* OR PT clinical trial

**Supplemental Table 5:** Reasons for excluding studies following full text analysis\*

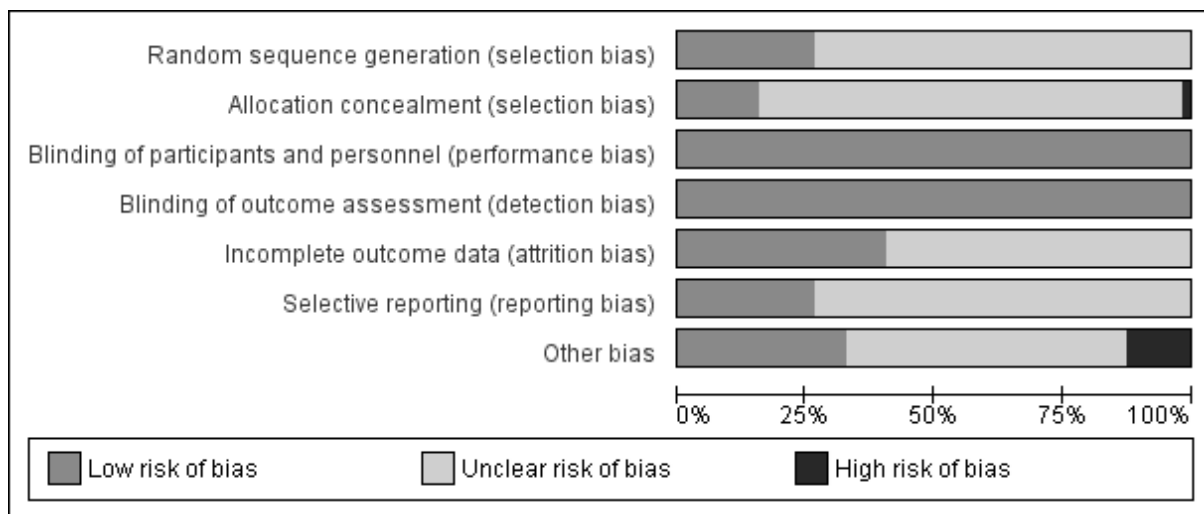
Study Citation	Reason for exclusion
Nil author 2013 (1)	Not RCT
Alfa 2017 (2)	Duplicate
Azcarate-Peril 2013 (3)	Not healthy study population
Azcarate-Peril 2016 (4)	Not healthy study population
Azcarate-Peril 2017 (5)	Not healthy study population
Azpiroz 2016 (6)	Not healthy study population
Baer 2009 (7)	Abstracts or unpublished studies
Benus 2010 (8)	Non-fiber or multifactorial intervention
Brahe 2014 (9)	Duplicate
Breinholt 2005 (10)	Non-fiber or multifactorial intervention
Brighenti 1999 (11)	Not RCT
Casellas 2007 (12)	Not healthy study population
Chen 2006 (13)	Not RCT
Chen 2008 (14)	Not healthy study population
Christensen 2013 (15)	Non-fiber or multifactorial intervention
Chung 2007 (16)	Not RCT
Clarke 2016 (17)	Duplicate
Clarke 2016 (18)	Duplicate
Clarke 2016 (19)	Duplicate
Cooper 2016 (20)	Abstracts or unpublished studies
Costabile 2016 (21)	Not RCT
Culpepper 2012 (22)	Abstracts or unpublished studies
Davis 2010 (23)	Not RCT
Davis 2011 (24)	Not RCT
De Preter 2007 (25)	Not RCT
Demircioglu 2008 (26)	Non-fiber or multifactorial intervention
Dewulf 2011 (27)	Abstracts or unpublished studies
Dewulf 2012 (28)	Abstracts or unpublished studies
Eastwood 1995 (29)	Non-fiber or multifactorial intervention
Eid 2015 (30)	Non-fiber or multifactorial intervention
Elison 2016 (31)	Duplicate
Famdodu 2016 (32)	Abstracts or unpublished studies
Famodu 2016 (33)	Abstracts or unpublished studies
Fava 2013 (34)	Non-fiber or multifactorial intervention
Finley 2007 (35)	Did not report on review outcomes
Ford 2017 (36)	Abstracts or unpublished studies
Gopal 2003 (37)	Duplicate
Gordon 2017 (38)	Abstracts or unpublished studies
Grasten 2000 (39)	Did not report on review outcomes
Guetterman 2016 (40)	Non-fiber or multifactorial intervention
Guglielmetti 2013 (41)	Non-fiber or multifactorial intervention
Hald 2016 (42)	Not healthy study population
Halmos 2013 (43)	Duplicate
Halmos 2014 (44)	Not healthy study population
Halmos 2015 (45)	Duplicate
Healey 2016 (46)	Abstracts or unpublished studies
Heiman 2014 (47)	Not healthy study population
Holscher 2014 (48)	Duplicate
Holscher 2015 (49)	Abstracts or unpublished studies
Hooda 2012 (50)	Secondary publication
Jalanka 2016 (51)	Abstracts or unpublished studies
Jenkins 1999 (52)	Did not report on review outcomes
Karl 2017 (53)	Duplicate
Kellow 2014 (54)	Not healthy study population
Klinder 2016 (55)	Non-fiber or multifactorial intervention

Study Citation	Reason for exclusion
Klosterbuer 2013 (56)	Did not report on review outcomes
Kolida 2007 (57)	Not RCT
Kovatcheva-Datchary 2015 (58)	Did not report on review outcomes
Kruse 1999 (59)	Not RCT
Lambert 2014 (60)	Abstracts or unpublished studies
Lambert 2015 (61)	Not healthy study population
Lamichhane 2014 (62)	Did not report on review outcomes
Langlands 2004 (63)	Not RCT
Lappi 2013 (64)	Not healthy study population
Lee 2016 (65)	Did not report on review outcomes
Lehtinen 2012 (66)	Abstracts or unpublished studies
Li 2009 (67)	Non-fiber or multifactorial intervention
Li 2014 (68)	Abstracts or unpublished studies
Li 2015 (69)	Abstracts or unpublished studies
Lin 2014 (70)	Not healthy study population
Lin 2016 (71)	Duplicate
Linetzky 2012 (72)	Not healthy study population
Lomax 2012 (73)	Duplicate
Lomax 2013 (74)	Duplicate
Lomax 2013 (75)	Abstracts or unpublished studies
Mai 2009 (76)	Abstracts or unpublished studies
Mai 2012 (77)	Non-fiber or multifactorial intervention
Maki 2011 (78)	Abstracts or unpublished studies
Marteau 2011 (79)	Not healthy study population
Matthan 2015 (80)	Abstracts or unpublished studies
Mayengbam 2017 (81)	Abstracts or unpublished studies
Medina-Vera 2017 (82)	Abstracts or unpublished studies
Mego 2017 (83)	Non-fiber or multifactorial intervention
Mitchell 2015 (84)	Not healthy study population
Mitsou 2009 (85)	Non-fiber or multifactorial intervention
Mitsou 2011 (86)	Non-fiber or multifactorial intervention
Orrhage 2000 (87)	Non-fiber or multifactorial intervention
Pantophlet 2017 (88)	Not RCT
Ramirez-Farias 2009 (89)	Secondary publication
Ramprasath 2015 (90)	Abstracts or unpublished studies
Rao 2001 (91)	Not RCT
Ravn-Haren 2013 (92)	Non-fiber or multifactorial intervention
Robinson 2001 (93)	Not RCT
Salazar 2013 (94)	Abstracts or unpublished studies
Salazar 2015 (95)	Abstracts or unpublished studies
Salden 2015 (96)	Abstracts or unpublished studies
Salonen 2014 (97)	Not healthy study population
Scarpellini 2012 (98)	Abstracts or unpublished studies
Scarpellini 2016 (99)	Did not report on review outcomes
Scholtens 2006 (100)	Did not report on review outcomes
Sloan 2016 (101)	Abstracts or unpublished studies
Smilowitz 2017 (102)	Not RCT
Song 2015 (103)	Non-fiber or multifactorial intervention
Souza 2015 (104)	Not healthy study population
Surakka 2009 (105)	Not healthy study population
Tannock 2004 (106)	Not RCT
Taylor 2016 (107)	Non-fiber or multifactorial intervention
Thompson 2016 (108)	Abstracts or unpublished studies
Thompson 2016 (109)	Abstracts or unpublished studies
Tomono 2010 (110)	Not healthy study population
Tuohy 2001 (111)	Not RCT
Tuohy 2001 (112)	Duplicate

<b>Study Citation</b>	<b>Reason for exclusion</b>
Ukhanova 2014 (113)	Non-fiber or multifactorial intervention
Upadhyaya 2016 (114)	Not healthy study population
Vanegas 2016 (115)	Abstracts or unpublished studies
Vanegas 2017 (116)	Secondary publication
Vanegas 2017 (117)	Duplicate
Vendrame 2011 (118)	Non-fiber or multifactorial intervention
Venkataraman 2016 (119)	Not RCT
Vitaglione 2015 (120)	Non-fiber or multifactorial intervention
Vulevic 2013 (121)	Not healthy study population
Walker 2011 (122)	Not healthy study population
Wallace 2015 (123)	Not RCT
Weickert 2011 (124)	Not healthy study population
West 2012 (125)	Not RCT
Westreich 2017 (126)	Abstracts or unpublished studies
Whisner 2016 (127)	Not healthy study population
Willis 2013 (128)	Did not report on review outcomes
Windey 2015 (129)	Did not report on review outcomes
Wong 2010 (130)	Not RCT
Wood 2017 (131)	Abstracts or unpublished studies
Wood 2017 (132)	Abstracts or unpublished studies
Worthley 2009 (133)	Not RCT
Worthley 2009 (134)	Abstracts or unpublished studies
Wutzke 2012 (135)	Abstracts or unpublished studies
Xiao 2014 (136)	Not RCT
Yang 2015 (137)	Not healthy study population
Yen 2011 (138)	Duplicate
Yen 2011 (139)	Not healthy study population
Yen 2011 (140)	Not healthy study population

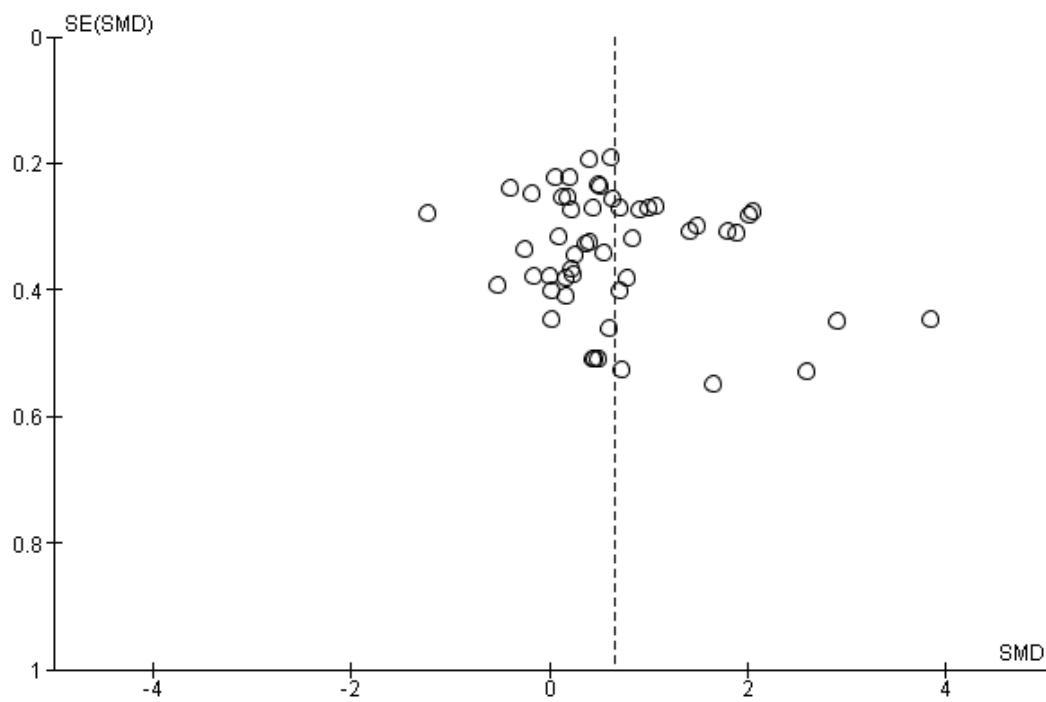
\* Citation numbers do not correspond to citations in main manuscript, and are provided at the end of this document.

## Risk of Bias



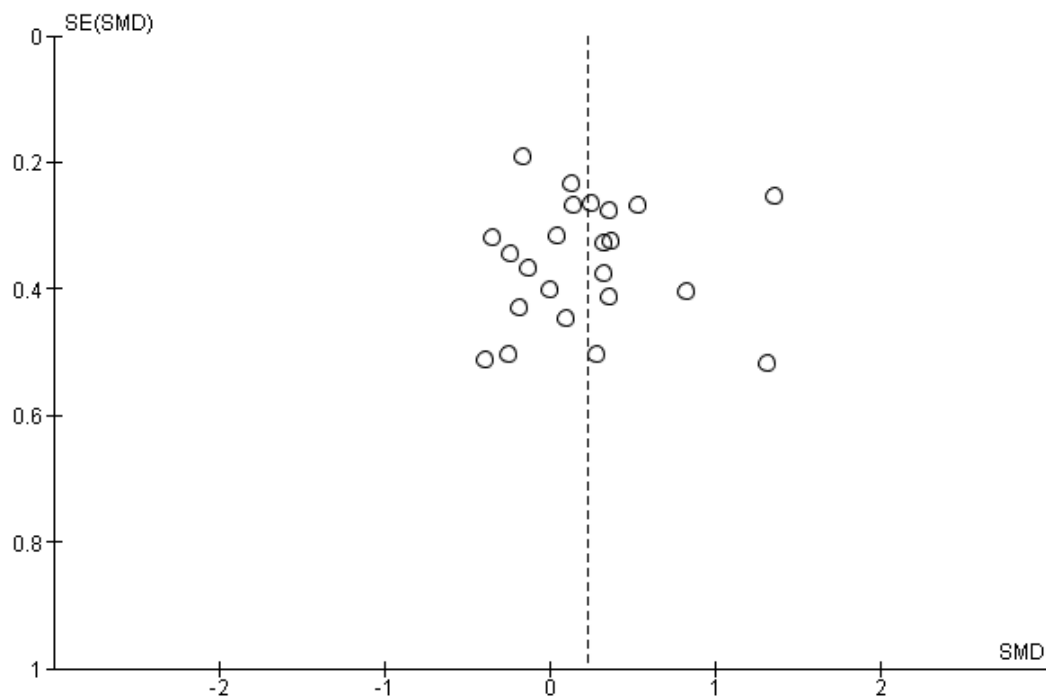
**Supplemental Figure 1:** Risk of bias across the included studies showing the summary percentage in each domain

## Reporting Bias

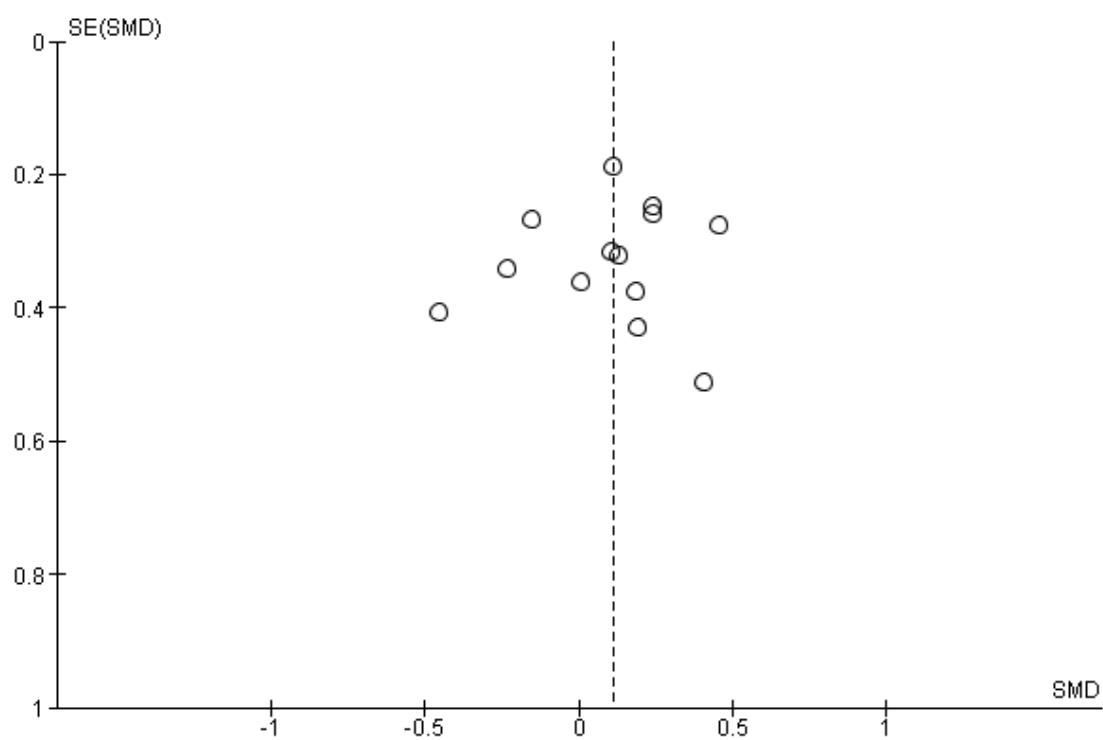


**Supplemental Figure 2:** Funnel plot for the effect of dietary fiber on *Bifidobacterium* spp. abundance

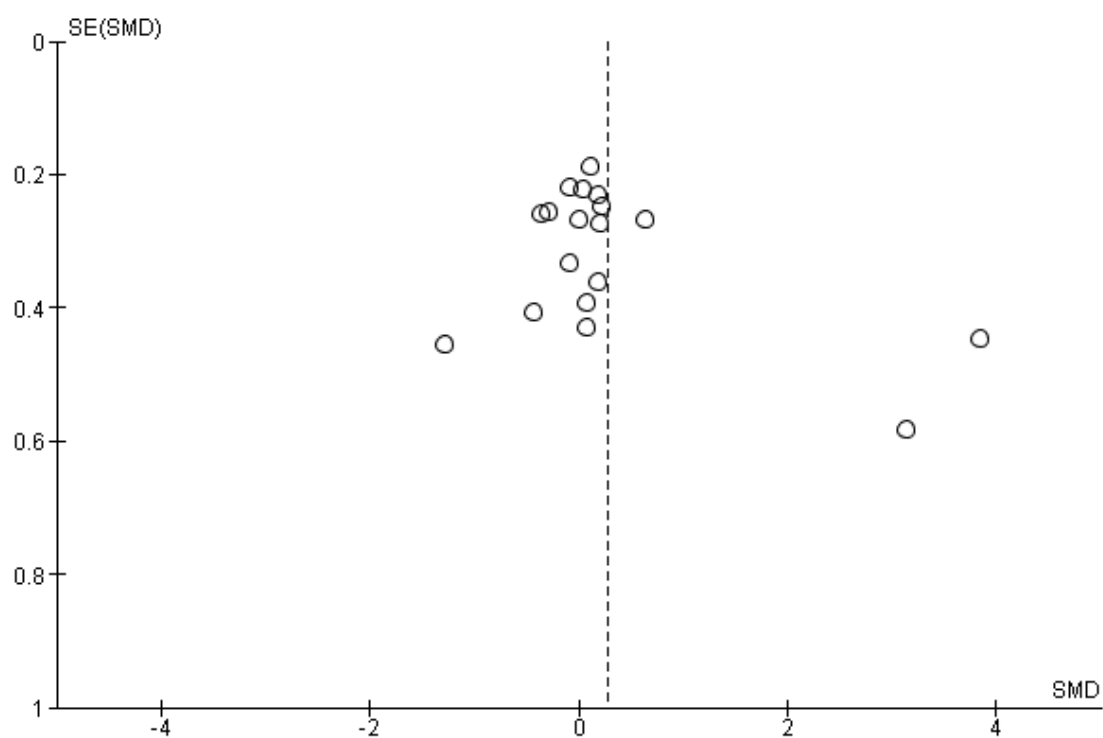




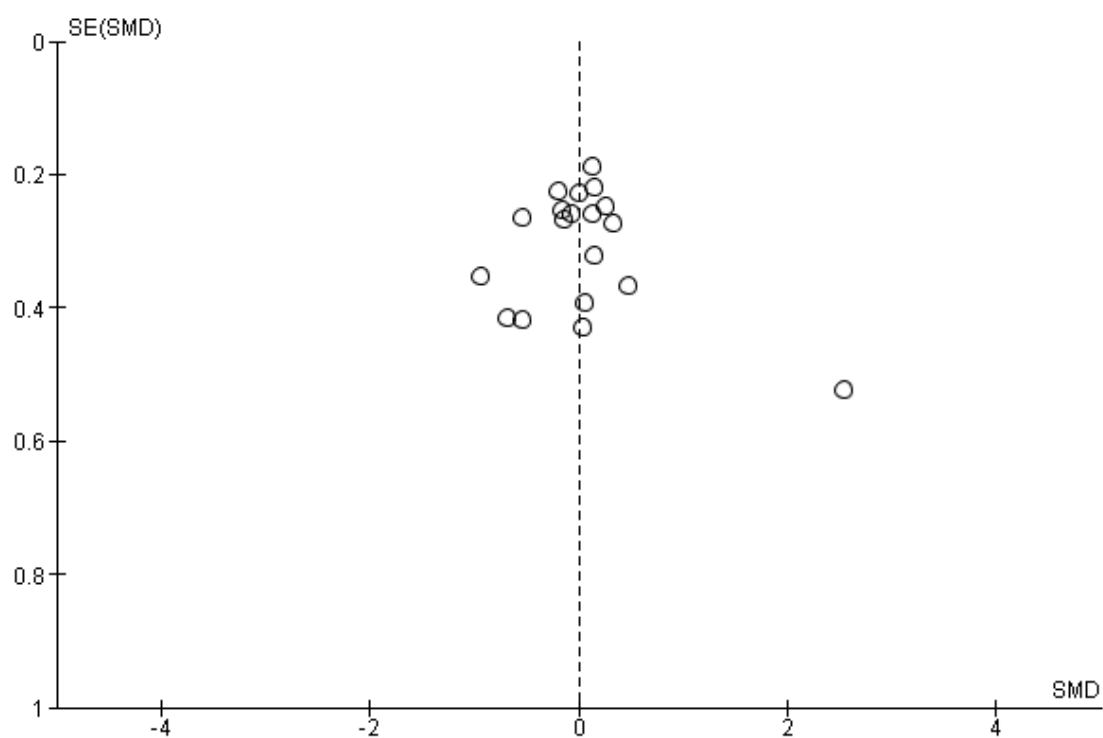
**Supplemental Figure 3:** Funnel plot for the effect of dietary fiber on *Lactobacillus* spp. abundance



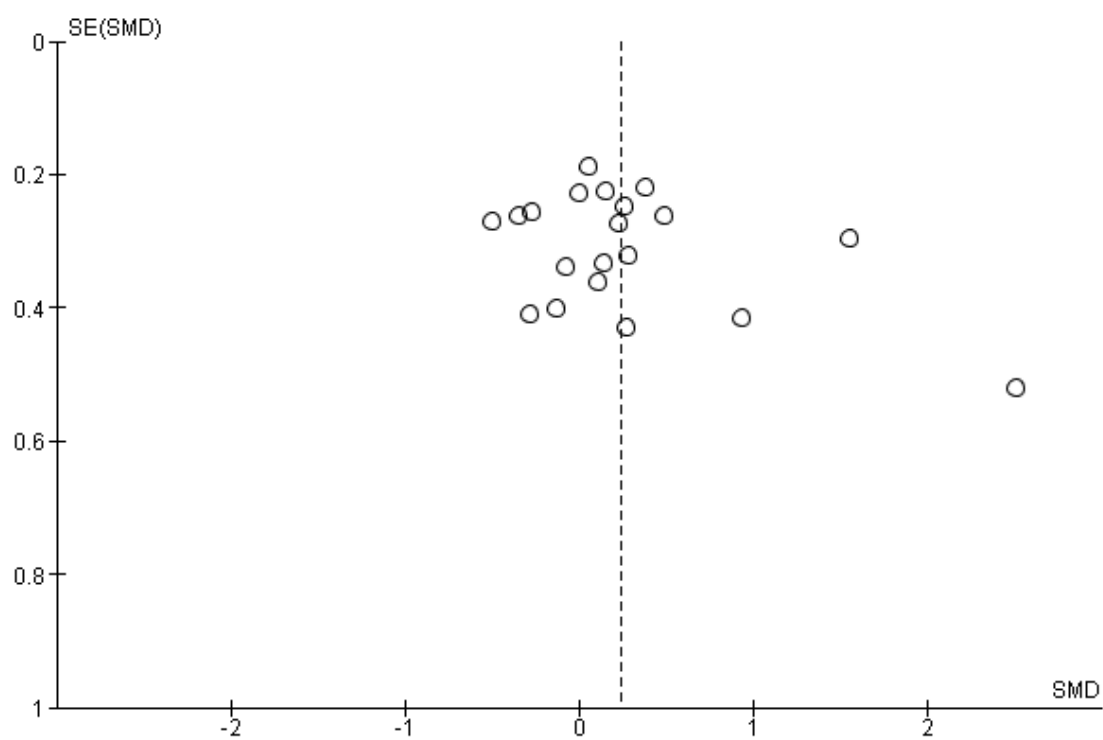
**Supplemental Figure 4:** Funnel plot for the effect of dietary fiber on total fecal SCFA



**Supplemental Figure 5:** Funnel plot for the effect of dietary fiber on fecal acetate



**Supplemental Figure 6:** Funnel plot for the effect of dietary fiber on fecal propionate



**Supplemental Figure 7:** Funnel plot for the effect of dietary fiber on fecal butyrate

**Supplemental Table 6:** Outcomes of pre-defined subgroup analyses undertaken

Outcome	Subgroup analysis	Subgroup difference (I <sup>2</sup> )	Subgroups	Studies in subgroup (n)	Result	P	Heterogeneity		
					Meta-analysis overall estimate (95% CI)		Chi-squared test	P	I <sup>2</sup>
Shannon Diversity Index	Trial design	0%	Cross-over	3	MD: -0.10 (95% CI: -0.19, -0.01)	0.03	1.36	0.51	0%
			Parallel	3	MD: -0.03 (95% CI: -0.57, 0.51)	0.91	9.35	0.009	79%
<i>Bifidobacterium</i> spp.	Intervention type	68.6%	Food	10	SMD: 0.75 (95% CI: 0.52, 0.98)	<0.00001	234.35	<0.00001	83%
			Supplement	41	SMD: 0.20 (95% CI: -0.36, 0.76)	0.49	76.94	<0.00001	88%
	Fiber type	45.3%	Accepted prebiotic	23	SMD: 0.68 (95% CI: 0.38, 0.98)	<0.00001	117.8	<0.00001	81%
			Candidate prebiotic	13	SMD: 0.77 (95% CI: 0.30, 1.24)	0.001	86.19	<0.00001	86%
			General fiber	14	SMD: 0.25 (95% CI: -0.16, 0.65)	0.24	80.54	<0.00001	84%
	Dose response	8.8%	≤5g/d	11	SMD: 0.51 (95% CI: 0.18, 0.84)	0.003	33.52	0.0002	70%
			5-10g/d	18	SMD: 0.48 (95% CI: 0.13, 0.84)	0.007	133.22	<0.00001	87%
			>10g/d	22	SMD: 0.85 (95% CI: 0.45, 1.25)	<0.00001	143.72	<0.00001	85%
	Trial design	77%	Cross-over	30	SMD: 0.44 (95% CI: 0.21, 0.66)	<0.00001	149.67	<0.00001	81%
			Parallel	21	SMD: 0.98 (95% CI: 0.52, 1.44)	<0.00001	148.63	<0.00001	87%
	Analysis method	0%	Culture	13	SMD: 0.70 (95% CI: 0.07, 1.33)	0.03	99.72	<0.00001	88%
			qPCR	11	SMD: 0.62 (95% CI: 0.29, 0.94)	0.0002	30.28	0.0008	67%
			FISH	19	SMD: 0.71 (95% CI: 0.31, 1.10)	0.0004	187.79	<0.00001	90%
			Sequencing	4	SMD: 0.61 (95% CI: 0.27, 0.95)	0.0005	0.83	0.84	0%
<i>Lactobacillus</i> spp.	Intervention type	0%	Food	4	SMD: 0.35 (95% CI: -0.46, 1.16)	0.40	18.73	0.00003	84%
			Supplement	19	SMD: 0.16 (95% CI: 0.01, 0.31)	0.04	19.27	0.38	7%
	Fiber type	69.1%	Accepted prebiotic	9	SMD: 0.34 (95% CI: 0.13, 0.55)	0.002	7.63	0.47	0%
			Candidate prebiotic	7	SMD: -0.06 (95% CI: -0.29, 0.16)	0.58	3.52	0.74	0%
			General fiber	7	SMD: 0.22 (95% CI: -0.31, 0.75)	0.42	23.23	0.0007	74%
	Dose	0%	≤5g/d	6	SMD: 0.16 (95% CI: -0.24, 0.56)	0.44	9.67	0.09	48%

Outcome	Subgroup analysis	Subgroup difference (I <sup>2</sup> )	Subgroups	Studies in subgroup (n)	Result	P	Heterogeneity		
					Meta-analysis overall estimate (95% CI)		Chi-squared test	P	I <sup>2</sup>
<i>Faecalibacterium prausnitzii</i>	response		5-10g/d	5	SMD: 0.14 (95% CI: -0.12, 0.39)	0.29	3.23	0.52	0%
			>10g/d	12	SMD: 0.29 (95% CI: -0.01, 0.59)	0.06	26.08	0.006	58%
			Cross-over	11	SMD: 0.08 (95% CI: -0.09, 0.25)	0.38	9.04	0.53	0%
	Trial design	57.7%	Parallel	12	SMD: 0.37 (95% CI: 0.04, 0.70)	0.03	26.8	0.005	59%
			Culture	7	SMD: 0.61 (95% CI: 0.13, 1.08)	0.01	15.99	0.01	62%
	Analysis method	55.1%	qPCR	9	SMD: 0.13 (95% CI: -0.07, 0.33)	0.21	7.36	0.50	0%
			FISH	2	SMD: -0.15 (95% CI: -0.48, 0.18)	0.37	0.01	0.94	0%
			Sequencing	3	SMD: 0.18 (95% CI: -0.19, 0.56)	0.33	1.53	0.46	0%
			≤5g/d	3	SMD: -0.10 (95% CI: -0.39, 0.19)	0.51	2.71	0.26	26%
	Dose response	38.0%	5-10g/d	6	SMD: -0.05 (95% CI: -0.23, 0.13)	0.57	2.55	0.77	0%
			>10g/d	4	SMD: 0.39 (95% CI: -0.09, 0.87)	0.11	6.24	0.10	52%
			Cross-over	8	SMD: 0.06 (95% CI: -0.18, 0.29)	0.63	12.71	0.08	45%
	Trial design	53.6%	Parallel	5	SMD: 0.60 (95% CI: -0.09, 1.29)	0.009	22.6	0.0002	82%
			Cross-over	2	SMD: -0.09 (95% CI: -0.46, 0.29)	0.65	0.25	0.62	0%
<i>Roseburia</i> spp.	Trial design	89.2%	Parallel	2	SMD: 0.71 (95% CI: 0.36, 1.06)	<0.00001	0.64	0.42	0%

**Supplemental Table 7:** Outcomes of post hoc subgroup analyses undertaken

Outcome	Subgroup analysis	Subgroup difference (I <sup>2</sup> )	Subgroups	Studies in subgroup (n)	Result	Heterogeneity			
					Meta-analysis overall estimate (95% CI)	P	Chi-squared test	P	I <sup>2</sup>
Total SCFA	Reporting method	44.5%	Dry weight of feces	6	SMD: 0.02 (95% CI: -0.23, 0.26)	0.89	2.81	0.73	0%
			Wet weight of feces	6	SMD: 0.25 (95% CI: 0.01, 0.49)	0.04	0.80	0.98	0%
Acetate	Reporting method	77.3%	Dry weight of feces	6	SMD: -0.08 (95% CI: -0.40, 0.25)	0.65	10.26	0.07	51%
			Wet weight of feces	10	SMD: 0.69 (95% CI: 0.05, 1.33)	0.03	98.97	<0.00001	91%
Propionate	Reporting method	0%	Dry weight of feces	6	SMD: -0.07 (95% CI: -0.33, 0.20)	0.61	7.15	0.21	30%
			Wet weight of feces	11	SMD: 0.09 (95% CI: -0.26, 0.44)	0.61	38.22	<0.00001	74%
Butyrate	Reporting method	74.1%	Dry weight of feces	7	SMD: 0.02 (95% CI: -0.18, 0.22)	0.81	1.26	0.97	0%
			Wet weight of feces	11	SMD: 0.47 (95% CI: 0.07, 0.87)	0.02	49.36	<0.00001	80%



## References to Table 5 citations

1. Prebiotic treatment of metabolic syndrome to reduce disease risk. 2013;0:24-6.
2. Alfa M, Strang D, Tappia P, Graham M, Domselaar G, Forbes J, Laminman V, Olson N, DeGagne P, Bray D, et al. A randomized trial to determine the impact of a digestion resistant starch composition on the gut microbiome in older and mid-age adults. 2017. Internet: <http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/220/CN-01366220/frame.html> accessed [Date Accessed]|.
3. Azcarate-Peril MA, Savaiano DA, Ritter AJ, Klaenhammer T. Microbiome alterations of lactose intolerant individuals in response to dietary intervention with galacto-oligosaccharides may help negate symptoms of lactose intolerance. *Gastroenterology* 2013;144(5):S893.
4. Azcarate-Peril MA, Ritter A, Savaiano D, Klaenhammer T. Impact of short-chain galactooligosaccharides on the gut microbiome of lactose intolerant individuals. *Gastroenterology* 2016;150(4):S424.
5. Azcarate-Peril MA, Ritter AJ, Savaiano D, Monteagudo-Mera A, Anderson C, Magness ST, Klaenhammer TR. Impact of short-chain galactooligosaccharides on the gut microbiome of lactose-intolerant individuals. *Proceedings of the National Academy of Sciences of the United States of America* 2017;114(3):E367-E75.
6. Azpiroz F, Molne L, Mendez S, Nieto A, Manichanh C, Mego M, Accarino A, Santos J, Sailer M, Theis S, et al. Effect of Chicory-derived Inulin on Abdominal Sensations and Bowel Motor Function. *Journal of Clinical Gastroenterology* 2016;0.
7. Baer DJ, Mai V, Okuma K, Tagami H, Kanahori S, Henderson T, Stote KS, Paul DR, Gordon DT, Rumpler WV. Metabolizable energy value of resistant maltodextrin. *The FASEB Journal* 2009;23.
8. Benus R, Werf T, Welling G, Judd P, Taylor M, Harmsen H, Whelan K. Association between *Faecalibacterium prausnitzii* and dietary fiber in colonic fermentation in healthy human subjects. *The British journal of nutrition* 2010;104(5):693-700.
9. Brahe L, Le Chatelier E, Prifti E, Kennedy S, Blädel T, Ha'kansson J, Pedersen O, Astrup A, Ehrlich S, Larsen L. Dietary intervention modulates the gut microbiota and improves insulin resistance-a randomized controlled trial in obese postmenopausal women. *Obesity Reviews* 2014;15:41-2.
10. Brejnholt SM, Tannock GW, Moller PL, Munro K, Tetens I. A rye bran diet, rich in plant lignans, has no influence on the composition of the gut microflora in postmenopausal women. *Microbial Ecology in Health and Disease* 2005;17(1):21-7.
11. Brighenti F, Casiraghi MC, Canzi E, Ferrari A. Effect of consumption of a ready-to-eat breakfast cereal containing inulin on the intestinal milieu and blood lipids in healthy male volunteers. *European Journal of Clinical Nutrition* 1999;53(9):726-33.
12. Casellas F, Borruel N, Torrejón A, Varela E, Antolin M, Guarner F, Malagelada JR. Oral oligofructose-enriched inulin supplementation in acute ulcerative colitis is well tolerated and associated with lowered faecal calprotectin. *Alimentary Pharmacology and Therapeutics* 2007;25(9):1061-7.
13. Chen HL, Cheng HC, Liu YJ, Liu SY, Wu WT. Konjac acts as a natural laxative by increasing stool bulk and improving colonic ecology in healthy adults. *Nutrition* 2006;22(11):1112-9.
14. Chen HL, Cheng HC, Wu WT, Liu YJ, Liu SY. Supplementation of konjac glucomannan into a low-fiber Chinese diet promoted bowel movement and improved colonic ecology in constipated adults: A placebo-controlled, diet-controlled trial. *Journal of the American College of Nutrition* 2008;27(1):102-8.
15. Christensen EG, Licht TR, Kristensen M, Bahl MI. Bifidogenic effect of whole-grain wheat during a 12-week energy-restricted dietary intervention in postmenopausal women. *European Journal of Clinical Nutrition* 2013;67(12):1316-21.
16. Chung YC, Hsu CK, Ko CY, Chan YC. Dietary intake of xylooligosaccharides improves the intestinal microbiota, fecal moisture, and pH value in the elderly. *Nutrition Research* 2007;27(12):756-61.
17. Clarke ST, Green-Johnson JM, Brooks SPJ, Ramdath DD, Bercik P, Avila C, Inglis GD, Green J, Yanke LJ, Selinger LB, et al.  $\beta$ -2-1 Fructan supplementation alters host immune responses in a manner consistent with increased exposure to microbial components: Results from a double-blinded, randomised, cross-over study in healthy adults. *British Journal of Nutrition* 2016;115(10):1748-59.
18. Clarke S, Green-Johnson J, Brooks S, Ramdath D, Bercik P, Avila C, Inglis G, Green J, Yanke L, Selinger L, et al. 2016;115:1748-59. Internet: <http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/551/CN-01179551/frame.html> accessed [Date Accessed]|.

19. Clarke S, T. r, Green-Johnson JM, Brooks SPJ, Ramdath DD, Bercik P, Avila C, Inglis GD, Green J, Yanke LJ, et al.  $\beta$ -2-1 Fructan supplementation alters host immune responses in a manner consistent with increased exposure to microbial components: results from a double-blinded, randomised, cross-over study in healthy adults. *British Journal of Nutrition* 2016;115(10):1748-59.
20. Cooper D, Kim EB, Marco M, Rust B, Welch L, Horn W, Martin R, Keim N. Relationship between human Gut microbiota and interleukin 6 levels in overweight and obese adults. *FASEB Journal* 2016;30.
21. Costabile A, Deaville E, Morales A, Gibson G. Prebiotic Potential of a Maize-Based Soluble Fiber and Impact of Dose on the Human Gut Microbiota. 2016;11:e0144457. Internet: <http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/505/CN-01168505/frame.html> accessed (Date Accessed)].
22. Culpepper T, Girard SA, Dahl W, Langkamp-Henken B, Mai V. Effects of galactooligosaccharides (GOS) on the gut microbiota of aged adults. *FASEB Journal* 2012;26.
23. Davis LMG, Martínez I, Walter J, Hutkins R. A dose dependent impact of prebiotic galactooligosaccharides on the intestinal microbiota of healthy adults. *International Journal of Food Microbiology* 2010;144(2):285-92.
24. Davis LM, Martinez I, Walter J, Goin C, Hutkins RW. Barcoded pyrosequencing reveals that consumption of galactooligosaccharides results in a highly specific bifidogenic response in humans. *PLoS ONE [Electronic Resource]* 2011;6(9):e25200.
25. De Preter V, Vanhoutte T, Huys G, Swings J, De Vuyst L, Rutgeerts P, Verbeke K. Effects of *Lactobacillus casei* Shirota, *Bifidobacterium breve*, and oligofructose-enriched inulin on colonic nitrogen-protein metabolism in healthy humans. *American Journal of Physiology - Gastrointestinal and Liver Physiology* 2007;292(1):G358-G68.
26. Demircioğlu Y, Başoğlu S, Özkan S, Şimşek I, Abbasoğlu U. The prebiotic effects of mixed sweetener containing polydextrose and oligofructose substituted sugar in diet. *Turkish Journal of Pharmaceutical Sciences* 2008;5(2):95-106.
27. Dewulf E, Cani P, Claus S, Neyrinck A, Gibson G, Thissen J, Delzenne N. Prebiotic approach contributes to metabolism improvement in obese women by changing the gut microbiota composition. *Annals of Nutrition and Metabolism* 2011;58:34.
28. Dewulf E, Cani P, Claus S, Neyrinck A, Puylaert P, Glenn G, De Vos W, Thissen JP, Delzenne N. Inulin-type fructans with prebiotic properties lessen endotoxemia and modulate host metabolism by changing gut microbiota composition in obese women. *Obesity Facts* 2012;5:200-1.
29. Eastwood MA, Allgood GS. The effect of olestra on breath gas production and faecal microbial counts. *European Journal of Clinical Nutrition* 1995;49(9):627-39.
30. Eid N, Osmanova H, Natchez C, Walton G, Costabile A, Gibson G, Rowland I, Spencer JPE. Impact of palm date consumption on microbiota growth and large intestinal health: A randomised, controlled, cross-over, human intervention study. *British Journal of Nutrition* 2015;114(8):1226-36.
31. Elison E, Vigsnaes LK, Rindom Krogsgaard L, Rasmussen J, Sorensen N, McConnell B, Hennet T, Sommer MOA, Bytzer P. Oral supplementation of healthy adults with 2'-O-fucosyllactose and lacto-N-neotetraose is well tolerated and shifts the intestinal microbiota. *The British journal of nutrition* 2016;116(8):1356-68.
32. Famodu OA, Cuff CF, Cockburn A, Downes MT, Murray PJ, McFadden JW, Colby SE, Morrell JS, Olfert IM, Olfert MD. Impact of free-living nutrition intervention on microbiome in college students at risk for Disease: FRUVEDomic pilot study. *FASEB Journal* 2016;30.
33. Famodu O, Cuff C, Cockburn A, Downes M, Murray P, McFadden J, Colby S, Morrell J, Olfert I, Olfert M. Impact of free-living nutrition intervention on microbiome in college students at risk for Disease: fRUVEDomic pilot study. 2016;30. Internet: <http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/516/CN-01266516/frame.html> accessed (Date Accessed)].
34. Fava F, Gitau R, Griffin BA, Gibson GR, Tuohy KM, Lovegrove JA. The type and quantity of dietary fat and carbohydrate alter faecal microbiome and short-chain fatty acid excretion in a metabolic syndrome 'at-risk' population. *International Journal of Obesity* 2013;37(2):216-23.
35. Finley JW, Burrell JB, Reeves PG. Pinto bean consumption changes SCFA profiles in fecal fermentations, bacterial populations of the lower bowel, and lipid profiles in blood of humans. *Journal of Nutrition* 2007;137(11):2391-8.
36. Ford AL, Macpherson C, Girard SA, Tompkins TA, Tremblay J, Christman M, Dahl WJ. Effects of a high protein diet with and without a multi-strain probiotic and prebiotic on microbiota and gastrointestinal

- wellness in older women: A randomized, double-blind, placebocontrolled crossover study. *FASEB Journal* 2017;31(1).
37. Gopal P, Prasad J, Gill H. Effects of the consumption of *Bifidobacterium lactis* HN019 (DR10) and galacto-oligosaccharides on the microflora of the gastrointestinal tract in human subjects. *Nutrition research (New York, NY)* 2003;23(10):1313-28.
  38. Gordon DT, Baer DJ, Mai V. Dietary fiber's contribution to the energy needs of the microbiota. *FASEB Journal* 2017;31(1).
  39. Gråsten SM, Juntunen KS, Poutanen KS, Gylling HK, Miettinen TA, Mykkänen HM. Rye bread improves bowel function and decreases the concentrations of some compounds that are putative colon cancer risk markers in middle-aged women and men. *Journal of Nutrition* 2000;130(9):2215-21.
  40. Guetterman HM, Swanson KS, Novotny JA, Baer DJ, Holscher HD. Walnut consumption influences the human gut microbiome. *FASEB Journal* 2016;30.
  41. Guglielmetti S, Fracassetti D, Taverniti V, Bo C, Vendrame S, Klimis-Zacas D, Arioli S, Riso P, Porrini M. Differential modulation of human intestinal bifidobacterium populations after consumption of a wild blueberry (*Vaccinium angustifolium*) drink. *Journal of agricultural and food chemistry* 2013;61(34):8134-40.
  42. Hald S, Schioldan AG, Moore ME, Dige A, Lærke HN, Agnholt J, Knudsen KEB, Hermansen K, Marco ML, Gregersen S, et al. Effects of arabinoxylan and resistant starch on intestinal microbiota and short-chain fatty acids in subjects with metabolic syndrome: A randomised crossover study. *PLoS ONE* 2016;11(7).
  43. Halmos E, Christophersen C, Bird A, Shepherd S, Gibson P, Muir J. The low FODMAP diet alters the composition of the colonic microbiota compared to a typical Australian intake in patients with irritable bowel syndrome: A randomised controlled cross-over trial. *Journal of Gastroenterology and Hepatology* 2013;28:122-3.
  44. Halmos EP, Christophersen CT, Bird AR, Shepherd SJ, Gibson PR, Muir JG. Diets that differ in their FODMAP content alter the colonic luminal microenvironment. *Gut* 2014;64(1):93-100.
  45. Halmos E, Christophersen C, Bird A, Shepherd S, Gibson P, Muir J. Diets that differ in their FODMAP content alter the colonic luminal microenvironment. *Gut* 2015;64(1):93-100.
  46. Healey G, Brough L, Butts C, Murphy R, Whelan K, Coad J. Influence of habitual dietary fiber intake on the responsiveness of the gut microbiota to a prebiotic: Protocol for a randomised, double-blind, placebo-controlled, cross-over, single-centre study. *BMJ Open* 2016;6(9).
  47. Heiman ML, Burton JH, Deych E, Shannon WD, Greenway FL. Improved oral glucose tolerance in prediabetics and type 2 diabetics (T2D) in a pilot clinical trial testing a novel gastrointestinal (GI) microbiome modulator. *Endocrine Reviews* 2014;35.
  48. Holscher H, Caporaso J, Brulc J, Swanson K. Fiber supplementation influences the phylogenetic structure and functional capacity of the adult human intestinal microbiome. *FASEB Journal* 2014;28(1).
  49. Holscher HD, Gregory Caporaso J, Hooda S, Brulc JM, Fahey GC, Swanson KS. Fiber supplementation influences phylogenetic structure and functional capacity of the human intestinal microbiome: Follow-up of a randomized controlled trial. *American Journal of Clinical Nutrition* 2015;101(1):55-64.
  50. Hooda S, Vester Boler BM, Rossoni Sero MC, Brulc JM, Staeger MA, Boileau TW, Dowd SE, Fahey Jr GC, Swanson KS. 454 pyrosequencing reveals a shift in fecal microbiota of healthy adult men consuming polydextrose or soluble corn fiber. *Journal of Nutrition* 2012;142(7):1259-65.
  51. Jalanka J, Sloan T, Major G, Krishnasamy S, Pritchard S, Mulvenna C, Lomer M, G, P, Spiller R. Associations between microbiota, colonic volume and transit during a low fodmap diet. *Gut* 2016;65:A51-A2.
  52. Jenkins DJA, Kendall CWC, Vuksan V, Augustin LSA, Li YM, Lee B, Mehling CC, Parker T, Faulkner D, Seyler H, et al. The effect of wheat bran particle size on laxation and colonic fermentation. *Journal of the American College of Nutrition* 1999;18(4):339-45.
  53. Philip Karl J, Meydani M, Barnett JB, Vanegas SM, Goldin B, Kane A, Rasmussen H, Saltzman E, Vangay P, Knights D, et al. Substituting whole grains for refined grains in a 6-wk randomized trial favorably affects energy-balance metrics in healthy men and postmenopausal women1-3. *American Journal of Clinical* 2017;105(3):589-99.
  54. Kellow NJ, Coughlan MT, Savage GS, Reid CM. Effect of dietary prebiotic supplementation on advanced glycation, insulin resistance and inflammatory biomarkers in adults with pre-diabetes: A study protocol for a double-blind placebo-controlled randomised crossover clinical trial. *BMC Endocrine Disorders* 2014;14.
  55. Klinder A, Shen Q, Heppel S, Lovegrove J, R, I, Tuohy K. Impact of increasing fruit and vegetables and flavonoid intake on the human gut microbiota. 2016;7:1788-96. Internet:

<http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/442/CN-01264442/frame.html> accessed Date Accessed)].

56. Klosterbuer AS, Hullar MAJ, Li F, Traylor E, Lampe JW, Thomas W, Slavin JL. Gastrointestinal effects of resistant starch, soluble maize fiber and pullulan in healthy adults. *British Journal of Nutrition* 2013;110(6):1068-74.
57. Kolida S, Meyer D, Gibson GR. A double-blind placebo-controlled study to establish the bifidogenic dose of inulin in healthy humans. *European Journal of Clinical Nutrition* 2007;61(10):1189-95.
58. Kovatcheva-Datchary P, Nilsson A, Akrami R, Lee YS, De Vadder F, Arora T, Hallen A, Martens E, Björck I, Bäckhed F. Dietary Fiber-Induced Improvement in Glucose Metabolism Is Associated with Increased Abundance of *Prevotella*. *Cell Metabolism* 2015;22(6):971-82.
59. Kruse H, Kleessen B, Blaut M. Effects of inulin on faecal bifidobacteria in human subjects. *The British journal of nutrition* 1999;82(5):375-82.
60. Lambert JE, Parnell JA, Han J, Sturzenegger T, Paul HA, Vogel HJ, Reimer RA. Evaluation of yellow pea fiber supplementation on weight loss and the gut microbiota: A randomized controlled trial. *BMC Gastroenterology* 2014;14(1).
61. Lambert JE, Parnell JA, Eksteen B, Raman M, Bomhof MR, Rioux KP, Madsen KL, Reimer RA. Gut microbiota manipulation with prebiotics in patients with non-alcoholic fatty liver disease: A randomized controlled trial protocol. *BMC Gastroenterology* 2015;15(1).
62. Lamichhane S, Yde C, Forssten S, Ouwehand A, Saarinen M, Jensen H, Gibson G, Rastall R, Fava F, Bertram H. Impact of dietary polydextrose fiber on the human gut metabolome. *Journal of agricultural and food chemistry* 2014;62(40):9944-51.
63. Langlands S, Hopkins M, Coleman N, Cummings J. Prebiotic carbohydrates modify the mucosa associated microflora of the human large bowel. *Gut* 2004;53(11):1610-6.
64. Lappi J, Salojärvi J, Kolehmainen M, Mykkänen H, Poutanen K, de Vos WM, Salonen A. Intake of whole-grain and fiber-rich rye bread versus refined wheat bread does not differentiate intestinal microbiota composition in finnish adults with metabolic syndrome. *Journal of Nutrition* 2013;143(5):648-55.
65. Lee I, Shi L, Webb D-L, Hellstrom PM, Riserus U, L, berg R. Effects of whole-grain rye porridge with added inulin and wheat gluten on appetite, gut fermentation and postprandial glucose metabolism: a randomised, cross-over, breakfast study. *The British journal of nutrition* 2016;116(12):2139-49.
66. Lehtinen MJ, Maneraat S, Childs CE, Forssten SD, Alhoniemi E, Yaqoob P, Ouwehand AC, Rastall RA. Consumption of *Bifidobacterium animalis* subsp. *Lactis* Bi-07 in a clinical trial enhances ex vivo phagocytic activity in healthy elderly adults. *Immunology* 2012;137:730.
67. Li F, Hullar MAJ, Schwarz Y, Lampe JW. Human gut bacterial communities are altered by addition of cruciferous vegetables to a controlled fruit- and vegetable-free diet. *Journal of Nutrition* 2009;139(9):1685-91.
68. Li Z, Finegold S, Summanen P, Downes J, Thames G, Corbett K, Dowd S, Krak M, Heber D. Xylooligosaccharide increases bifidobacteria but not lactobacilli demonstrating potential for obesity prevention and treatment. *FASEB Journal* 2014;28(1).
69. Li Z, Yang J, Carlsen P, Henning S, Hsu M, Tseng CH, Thames G, Finegold S, Heber D. Xylooligosaccharide induced changes in gut microbiota in healthy and prediabetic adults. *FASEB Journal* 2015;29(1).
70. Lin X, Wenkui Y, Jun J, Ning L. Clinical benefits after soluble dietary fiber supplementation; A randomized clinical trial in adults with slow-transit constipation. *National Medical Journal of China* 2014;94(48):3813-6.
71. Lin SH, Chou LM, Chien YW, Chang JS, Lin CI. Prebiotic Effects of Xylooligosaccharides on the Improvement of Microbiota Balance in Human Subjects. *Gastroenterology Research and Practice* 2016;2016.
72. Linetzky WD, Alves PC, Logullo L, Manzoni JT, Almeida D, Teixeira dSM, Matos dMTR. Microbiota benefits after inulin and partially hydrolyzed guar gum supplementation: a randomized clinical trial in constipated women. *Nutrición hospitalaria* 2012;27(1):123-9.
73. Lomax A, Cheung L, Tuohy K, Noakes P, Miles E, Calder P. beta2-1 Fructans have a bifidogenic effect in healthy middle-aged human subjects but do not alter immune responses examined in the absence of an in vivo immune challenge: Results from a randomised controlled trial. *British journal of nutrition* 2012;108(10):1818-28.
74. Lomax A, Noakes P, Miles E, Cheung L, Tuohy K, Calder P. beta2-1 fructans have a bifidogenic effect in healthy middle-aged humans and enhance the antibody response to seasonal influenza vaccination, but do not alter immune responses examined in the absence of vaccination: Results from a randomised controlled trial. *Proceedings of the Nutrition Society* 2013;72:E12.

75. Lomax AR, Noakes PS, Miles EA, Cheung L, Tuohy KM, Calder PC.  $\beta$ 2-1 fructans have a bifidogenic effect in healthy middle-aged humans and enhance the antibody response to seasonal influenza vaccination, but do not alter immune responses examined in the absence of vaccination: Results from a randomised controlled trial. *Proceedings of the Nutrition Society* 2013;72:E12.
76. Mai V, Ukhanova M, Baer D, Okuma K, Tagami H, Kanahori S, Henderson T, Gordon DT. Effects of resistant maltodextrin on fecal microbiota composition. *The FASEB Journal* 2009;23.
77. Mai V, Fredborg M, Ukhanova M, Wang X, Daniel S, Novotny J, Gebauer S, Baer D. Human gut microbiota changes after consumption of almonds or pistachios. *FASEB Journal* 2012;26.
78. Maki KC, Gibson G, Dickman R, Kendall CWC, Chen CYO, Almeida N, Blumberg J. A double-blind, randomized, controlled crossover trial to assess the prebiotic effects of arabinoxylan-oligosaccharides (AXOS) in healthy men and women. *FASEB Journal* 2011;25.
79. Marteau P, Jacobs H, Cazaubiel M, Signoret C, Prevel J, Housez B. Effects of chicory inulin in constipated elderly people: a double-blind controlled trial. *International journal of food sciences and nutrition* 2011;62(2):164-70.
80. Matthan NR, Kane AV, Johnson WE, Manimaran S, Faits T, Lichtenstein AH. Dietary carbohydrate quality affects plasma lipid profile and the microbiome. *Circulation* 2015;132.
81. Mayengbam S, Lambert JE, Parnell JA, Tunnicliffe JM, Han J, Sturzenegger T, Vogel HJ, Shearer J, Reimer RA. Dietary fiber supplementation normalizes Serum metabolites of adults with overweight/obesity in a 12-week randomized control trial. *FASEB Journal* 2017;31(1).
82. Medina-Vera I, Sanchez-Tood M, Aguilar-López M, Guevara-Cruz M, Flores-López A, Tovar AR, Torres N. Effect of a combination of functional foods (nopai, oat, chia seed and inulin) on the gut microbiota of subjects with Type 2 diabetes. *FASEB Journal* 2017;31(1).
83. Mego M, Manichanh C, Accarino A, Campos D, Pozuelo M, Varela E, Vulevic J, Tzortzis G, Gibson G, Guarner F, et al. Metabolic adaptation of colonic microbiota to galactooligosaccharides: a proof-of-concept study. *2017;45:670-80.* Internet: <http://onlinelibrary.wiley.com/doi/cochrane/central/articles/595/CN-01329595/frame.html> [accessed Date Accessed)].
84. Mitchell CM, Davy BM, Halliday TM, Hulver MW, Neilson AP, Ponder MA, Davy KP. The effect of prebiotic supplementation with inulin on cardiometabolic health: Rationale, design, and methods of a controlled feeding efficacy trial in adults at risk of type 2 diabetes. *Contemporary Clinical Trials* 2015;45.
85. Mitsou EK, Kougia E, Nomikos T, Yannakoulia M, Mountzouris KC, Kyriacou A. Effect of banana consumption on faecal microbiota: A randomised, controlled trial. *Anaerobe* 2011;17(6):384-7.
86. Mitsou EK, Turunen K, Anapliotis P, Zisi D, Spiliotis V, Kyriacou A. Impact of a jelly containing short-chain fructo-oligosaccharides and Sideritis euboea extract on human faecal microbiota. *International Journal of Food Microbiology* 2009;135(2):112-7.
87. Orrhage K, Sjöstedt S, Nord CE. Effect of supplements with lactic acid bacteria and oligofructose on the intestinal microflora during administration of cefpodoxime proxetil. *Journal of Antimicrobial Chemotherapy* 2000;46(4):603-11.
88. Pantophlet AJ, Wopereis S, Eelderink C, Vonk RJ, Stroeve JH, Bijlsma S, van Stee L, Bobeldijk I, Priebe MG. Metabolic profiling reveals differences in plasma concentrations of arabinose and xylose after consumption of fiber-rich pasta and wheat bread with differential rates of systemic appearance of exogenous glucose in healthy men. *Journal of* 2017;147(2):152-60.
89. Ramirez-Farias C, Slezak K, Fuller Z, Duncan A, Holtrop G, Louis P. Effect of inulin on the human gut microbiota: Stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*. *British Journal of Nutrition* 2009;101(4):541-50.
90. Ramprasath V, Thandapilly S, Yang S, Abraham A, Jones P, Ames N. Effect of consuming novel foods consisting high oleic canola oil, barley beta-glucan, and DHA on cardiovascular disease risk in humans: The CONFIDENCE (Canola Oil and Fiber with DHA Enhanced) study - protocol for a randomized controlled trial. *Trials* 2015;16(1).
91. Rao VA. The prebiotic properties of oligofructose at low intake levels. *Nutrition Research* 2001;21(6):843-8.
92. Ravn-Haren G, Dragsted LO, Buch-Andersen T, Jensen EN, Jensen RI, Németh-Balogh M, Paulovicsová B, Bergström A, Wilcks A, Licht TR, et al. Intake of whole apples or clear apple juice has contrasting effects on plasma lipids in healthy volunteers. *European Journal of Nutrition* 2013;52(8):1875-89.
93. Robinson R, Feirtag J, Slavin J. Effects of dietary arabinogalactan on gastrointestinal and blood parameters in healthy human subjects. *Journal of the American College of Nutrition* 2001;20(4):279-85.

94. Salazar N, Dewulf EM, Neyrinck AM, Bindels LB, Cani PD, Mahillon J, Gueimonde M, De Los Reyes Gavilán CG, Thissen JP, Delzenne NM. Prebiotic effect of inulin type fructans: A focus on bifidobacterium populations and microbial related metabolites in obese individuals. *Annals of Nutrition and Metabolism* 2013;63:1696.
95. Salazar N, Dewulf EM, Neyrinck AM, Bindels LB, Cani PD, Mahillon J, de Vos WM, Thissen JP, Gueimonde M, de los Reyes-Gavilán CG, et al. Inulin-type fructans modulate intestinal Bifidobacterium species populations and decrease fecal short-chain fatty acids in obese women. *Clinical Nutrition* 2015;34(3):501-7.
96. Salden B, Troost F, Wilms E, Brüll F, Truchado P, Van De Wiele T, Possemiers S, Masclee A. Arabinoxylans show distinct prebiotic properties and may affect intestinal barrier function. *Gastroenterology* 2015;148(4):S197.
97. Salonen A, Lahti L, Salojärvi J, Holtrop G, Korpela K, Duncan S, Date P, Farquharson F, Johnstone A, Lobley G, et al. Impact of diet and individual variation on intestinal microbiota composition and fermentation products in obese men. *The ISME journal* 2014;8(11):2218-30.
98. Scarpellini E, Deloof E, Vos R, Verbeke K, Francois I, Delcour JA, Broekaert WF, Tack JF. The influence of acute colonic fermentation by arabinoxylan-oligosaccharide (AXOS) administration on gastric sensorimotor function and nutrient tolerance in man. *Gastroenterology* 2012;142(5):S309.
99. Scarpellini E, Deloof E, Vos R, Francois I, Delcour J, Broekaert W, Verbeke K, Tack J. The effect of arabinoxyloligosaccharides on gastric sensory-motor function and nutrient tolerance in man. 2016;28:1194-203. Internet: <http://onlinelibrary.wiley.com/doi/10.1111/1751-2533.12893> [accessed Date Accessed].
100. Scholtens PAMJ, Alles MS, Willemsen LEM, van den Braak C, Bindels JG, Boehm G, Govers MJAP. Dietary fructo-oligosaccharides in healthy adults do not negatively affect faecal cytotoxicity: A randomised, double-blind, placebo-controlled crossover trial. *British Journal of Nutrition* 2006;95(6):1143-9.
101. Sloan TJ, Jalanka J, Major GA, Krishnasamy S, Pritchard S, Lomer M, Gowland P, Spiller RC. Associations between microbiota, colonic volume and transit and the low fodmap diet with and without added oligofructose. *Gastroenterology* 2016;150(4):S82.
102. Smilowitz JT, Lemay DG, Kalanetra KM, Chin EL, Zivkovic AM, Breck MA, German JB, Mills DA, Slupsky C, Barile D. Tolerability and safety of the intake of bovine milk oligosaccharides extracted from cheese whey in healthy human adults. *Journal of Science* 2017.
103. Song MY, Wang JH, Eom T, Kim H. Schisandra chinensis fruit modulates the gut microbiota composition in association with metabolic markers in obese women: A randomized, double-blind placebo-controlled study. *Nutrition Research* 2015;35(8):655-63.
104. Souza LSAM, Rodrigues V, Araujo T, Oliveira T, Do CGPM, Luces FFC. Yacon-Based Product in the Modulation of Intestinal Constipation. *Journal of medicinal food* 2015;18(9):980-6.
105. Surakka A, Kajander K, Rajilić-Stojanović M, Karjalainen H, Hatakka K, Vapaatalo H, Zoetendal EG, De Vos WM, Korpela R, Tynkkynen S. Yoghurt containing galactooligosaccharides facilitates defecation among elderly subjects and selectively increases the number of Bifidobacteria. *International Journal of Probiotics and Prebiotics* 2009;4(1):65-74.
106. Tannock GW, Munro K, Bibiloni R, Simon MA, Hargreaves P, Gopal P, Harmsen H, Welling G. Impact of Consumption of Oligosaccharide-Containing Biscuits on the Fecal Microbiota of Humans. *Applied and Environmental Microbiology* 2004;70(4):2129-36.
107. Taylor AM, Swanson KS, Novotny JA, Baer DJ, Holscher HD. Impact of almond consumption on the composition of the gastrointestinal microbiota of healthy adult men and women. *FASEB Journal* 2016;30.
108. Thompson S, Swanson K, Novotny J, Baer D, Holscher H. Gastrointestinal microbial changes following whole grain barley and oat consumption in healthy men and women. 2016;30. Internet: <http://onlinelibrary.wiley.com/doi/10.1111/1751-2533.12893> [accessed Date Accessed].
109. Thompson SV, Swanson KS, Novotny JA, Baer DJ, Holscher HD. Gastrointestinal microbial changes following whole grain barley and oat consumption in healthy men and women. *FASEB Journal* 2016;30.
110. Tomono Y, Yamamoto T, Yamaguchi H. Effect of synthesized inulin on bowel habit and fecal microflora in healthy adults with low fecal frequency. *Japanese Pharmacology and Therapeutics* 2010;38(11):1031-40.

111. Tuohy KM, Finlay RK, Wynne AG, Gibson GR. A human volunteer study on the prebiotic effects of HP-inulin - Faecal bacteria enumerated using fluorescent In situ hybridisation (FISH). *Anaerobe* 2001;7(3):113-8.
112. Tuohy KM, Kolida S, Lustenberger AM, Gibson GR. The prebiotic effects of biscuits containing partially hydrolysed guar gum and fructo-oligosaccharides--a human volunteer study. *British Journal of Nutrition* 2001;86(3):341-8.
113. Ukhanova M, Wang X, Baer DJ, Novotny JA, Fredborg M, Mai V. Effects of almond and pistachio consumption on gut microbiota composition in a randomised cross-over human feeding study. *British Journal of Nutrition* 2014;111(12):2146-52.
114. Upadhyaya B, Juenemann R, McCormack L, Fardin-Kia AR, Clapper J, Nichenametla S, Specker B, Dey M. Prebiotic diet modulates gut microbial composition and metabolic functions in metabolic syndrome patients: Follow-up of a double blind, controlled, crossover intervention. *FASEB Journal* 2016;30.
115. Vanegas SM, Meydani M, Barnett JB, Kane A, Goldin B, Wu D, Karl JP, Brown C, Vangay P, Knights D, et al. Effect of a diet rich in whole grains on gut microbiota, and immune and inflammatory markers of healthy adults. *FASEB Journal* 2016;30.
116. Vanegas SM, Meydani M, Barnett JB, Goldin B, Kane A, Rasmussen H, Brown C, Vangay P, Knights D, Jonnalagadda S, et al. Substituting whole grains for refined grains in a 6-wk randomized trial has a modest effect on gut microbiota and immune and inflammatory markers of healthy adults. *American Journal of Clinical* 2017;105(3):635-50.
117. Vanegas SM, Meydani M, Barnett JB, Goldin B, Kane A, Rasmussen H, Brown C, Vangay P, Knights D, Jonnalagadda S, et al. Substituting whole grains for refined grains in a 6-wk randomized trial has a modest effect on gut microbiota and immune and inflammatory markers of healthy adults. *The American journal of clinical nutrition* 2017;105(3):635-50.
118. Vendrame S, Guglielmetti S, Riso P, Arioli S, Klimis-Zacas D, Porrini M. Six-week consumption of a wild blueberry powder drink increases Bifidobacteria in the human gut. *Journal of agricultural and food chemistry* 2011;59(24):12815-20.
119. Venkataraman A, Sieber JR, Schmidt AW, Waldron C, Theis KR, Schmidt TM. Variable responses of human microbiomes to dietary supplementation with resistant starch. 2016;4.
120. Vitaglione P, Mennella I, Ferracane R, Rivellese AA, Giacco R, Ercolini D, Gibbons SM, La Stora A, Gilbert JA, Jonnalagadda S, et al. Whole-grain wheat consumption reduces inflammation in a randomized controlled trial on overweight and obese subjects with unhealthy dietary and lifestyle behaviors: Role of polyphenols bound to cereal dietary fiber. *American Journal of Clinical Nutrition* 2015;101(2):251-61.
121. Vulevic J, Juric A, Tzortzis G, Gibson G. A mixture of trans-galactooligosaccharides reduces markers of metabolic syndrome and modulates the fecal microbiota and immune function of overweight adults. *The Journal of nutrition* 2013;143(3):324-31.
122. Walker AW, Ince J, Duncan SH, Webster LM, Holtrop G, Ze X, Brown D, Stares MD, Scott P, Bergerat A, et al. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *The Isme Journal* 2011;5(2):220-30.
123. Wallace AJ, Eady SL, Hunter DC, Skinner MA, Huffman L, Ansell J, Blatchford P, Wohlers M, Herath TD, Hedderley D, et al. No difference in fecal levels of bacteria or short chain fatty acids in humans, when consuming fruit juice beverages containing fruit fiber, fruit polyphenols, and their combination. *Nutrition Research* 2015;35(1):23-34.
124. Weickert MO, Arafat AM, Blaut M, Alpert C, Becker N, Leupelt V, Rudovich N, Möhlig M, Pfeiffer AF. Changes in dominant groups of the gut microbiota do not explain cereal-fiber induced improvement of whole-body insulin sensitivity. *Nutrition and Metabolism* 2011;8.
125. West NP, Pyne DB, Cripps AW, Christophersen CT, Conlon MA, Fricker PA. Gut balance, a synbiotic supplement, increases fecal *Lactobacillus paracasei* but has little effect on immunity in healthy physically active individuals. *Gut Microbes* 2012;3(3):1-7.
126. Westreich ST, Barile D, Salcedo J, Mills DA, Smilowitz JT, Korf I, Lemay DG. Using metatranscriptomics to determine effects of dietary supplementation with bovine milk oligosaccharides in healthy adults. *FASEB Journal* 2017;31(1).
127. Whisner C, Martin B, Nakatsu C, Story J, MacDonald-Clarke C, McCabe L, McCabe G, Weaver C. Soluble Corn Fiber Increases Calcium Absorption Associated with Shifts in the Gut Microbiome: a Randomized Dose-Response Trial in Free-Living Pubertal Females. 2016;146:1298-306. Internet: <http://onlinelibrary.wiley.com/doi/10.1111/ajcn.13333> accessed Date Accessed)].

128. Willis ND, Mann S, Xie L, McCallum IDJ, Kelly SB, Bradburn DM, Belshaw NJ, Johnson IT, Mathers JC. Impact of non-digestible carbohydrates on biomarkers of gastrointestinal health: A human intervention study. *Proceedings of the Nutrition Society* 2013;72:E260.
129. Windey K, De Preter V, Huys G, Broekaert WF, Delcour JA, Louat T, Herman J, Verbeke K. Wheat bran extract alters colonic fermentation and microbial composition, but does not affect faecal water toxicity: A randomised controlled trial in healthy subjects. *British Journal of Nutrition* 2015;113(2):225-38.
130. Wong JMW, Kendall CWC, De Souza R, Emam A, Marchie A, Vidgen E, Holmes C, Jenkins DJA. The effect on the blood lipid profile of soy foods combined with a prebiotic: A randomized controlled trial. *Metabolism: Clinical and Experimental* 2010;59(9):1331-40.
131. Wood LG, Berthon BS, Zapirain R, Leong LEX, Baines KJ, Gibson PG, Arnold D, Rogers GB. Asthma control, airway inflammation and gut microbiome are improved by soluble fiber supplementation. *Respirology* 2017;22:21.
132. Wood LG, Berthon BS, Zapirain R, Leong LEX, Baines KA, Gibson PG, Arnold D, Rogers G. Airway inflammation, asthma control and gut microbiome are improved by soluble fiber supplementation. *American Journal of Respiratory and Critical Care Medicine* 2017;195.
133. Worthley DL, Le Leu RK, Whitehall VL, Conlon M, Christophersen C, Belobrajdic D, Mallitt KA, Hu Y, Irahara N, Ogino S, et al. A human, double-blind, placebo-controlled, crossover trial of prebiotic, probiotic, and synbiotic supplementation: Effects on luminal, inflammatory, epigenetic, and epithelial biomarkers of colorectal cancer. *American Journal of Clinical Nutrition* 2009;90(3):578-86.
134. Worthley DL, Leleu R, Whitehall V, Conlon M, Christophersen C, Belobrajdic D, Mallitt K, Ogino S, Irahara N, Leggett B, et al. A human, double-blind, placebo-controlled, cross-over trial of prebiotic, probiotic and synbiotic supplementation: Effects on luminal, inflammatory, epigenetic and epithelial biomarkers of colorectal cancer. *Journal of Gastroenterology and Hepatology* 2009;24:A239.
135. Wutzke KD, Scholübbbers D. The metabolic effect of resistant starch and yoghurt on the colonic ammonia metabolism in humans as measured by lactose-[15N2]ureide. *Clinical Nutrition, Supplement* 2012;7(1):62-3.
136. Xiao S, Fei N, Pang X, Shen J, Wang L, Zhang B, Zhang M, Zhang X, Zhang C, Li M, et al. A gut microbiota-targeted dietary intervention for amelioration of chronic inflammation underlying metabolic syndrome. *FEMS Microbiology Ecology* 2014;87(2):357-67.
137. Yang J, Summanen PH, Henning SM, Hsu M, Lam H, Huang J, Tseng CH, Dowd SE, Finegold SM, Heber D, et al. Xylooligosaccharide supplementation alters gut bacteria in both healthy and prediabetic adults: A pilot study. *Frontiers in Physiology* 2015;6.
138. Yen C, Kuo Y, Tseng Y, Lee M, Chen H. Beneficial effects of fructo-oligosaccharides supplementation on fecal bifidobacteria and index of peroxidation status in constipated nursing-home residents--a placebo-controlled, diet-controlled trial. *Nutrition (Burbank, Los Angeles County, Calif)* 2011;27(3):323-8.
139. Yen C-H, Kuo Y-W, Tseng Y-H, Lee M-C, Chen H-L. Beneficial effects of fructo-oligosaccharides supplementation on fecal bifidobacteria and index of peroxidation status in constipated nursing-home residents—a placebo-controlled, diet-controlled trial. *Nutrition* 2011;27(3):323-8.
140. Yen CH, Tseng YH, Kuo YW, Lee MC, Chen HL. Long-term supplementation of isomalto-oligosaccharides improved colonic microflora profile, bowel function, and blood cholesterol levels in constipated elderly people-A placebo-controlled, diet-controlled trial. *Nutrition* 2011;27(4):445-50.